

PARENT COOPERATION TREATY

PCT

NOTIFICATION OF THE RECORDING
OF A CHANGE(PCT Rule 92bis.1 and
Administrative Instructions, Section 422)

Date of mailing (day/month/year) 07 December 2000 (07.12.00)
Applicant's or agent's file reference <u>41555/JMD</u>
International application No. PCT/GB00/00582

From the INTERNATIONAL BUREAU

To:

REDDIE & GROSE
16, Theobalds Road
London WC1X 8PL
ROYAUME-UNI

1. The following indications appeared on record concerning: <input type="checkbox"/> the applicant <input type="checkbox"/> the inventor <input checked="" type="checkbox"/> the agent <input type="checkbox"/> the common representative				
Name and Address HARRISON GODDARD FOOTE Tower House Merrion Way Leeds LS2 8PA United Kingdom	State of Nationality		State of Residence	
	Telephone No. +44 1223 360350			
	Facsimile No. +44 1223 360280			
	Teleprinter No.			
2. The International Bureau hereby notifies the applicant that the following change has been recorded concerning: <input type="checkbox"/> the person <input type="checkbox"/> the name <input type="checkbox"/> the address <input type="checkbox"/> the nationality <input type="checkbox"/> the residence				
Name and Address REDDIE & GROSE 16, Theobalds Road London WC1X 8PL United Kingdom	State of Nationality		State of Residence	
	Telephone No. +44 1223 360350			
	Facsimile No. +44 1223 360280			
	Teleprinter No.			
3. Further observations, if necessary:				
CORRECTED VERSION				
4. A copy of this notification has been sent to:				
<input checked="" type="checkbox"/> the receiving Office <input type="checkbox"/> the International Searching Authority <input checked="" type="checkbox"/> the International Preliminary Examining Authority		<input type="checkbox"/> the designated Offices concerned <input checked="" type="checkbox"/> the elected Offices concerned <input type="checkbox"/> other:		

The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland Facsimile No.: (41-22) 740.14.35	Authorized officer Dominique DELMAS Telephone No.: (41-22) 338.83.38
---	--

THIS PAGE BLANK (USPTO)

PARENT COOPERATION TREATY

PCT

NOTIFICATION OF THE RECORDING
OF A CHANGE(PCT Rule 92bis.1 and
Administrative Instructions, Section 422)Date of mailing (day/month/year)
21 February 2001 (21.02.01)

From the INTERNATIONAL BUREAU

To:

REDDIE & GROSE
Daedalus House
Station Road
Cambridge CB1 2RE
ROYAUME-UNIApplicant's or agent's file reference
41555/JMD

IMPORTANT NOTIFICATION

International application No.
PCT/GB00/00582International filing date (day/month/year)
18 February 2000 (18.02.00)

1. The following indications appeared on record concerning:

 the applicant the inventor the agent the common representative

Name and Address

HARRISON GODDARD FOOTE
Tower House
Merrion Way
Leeds LS2 8PA
United Kingdom

State of Nationality

State of Residence

Telephone No.

+44 113 290 1400

Facsimile No.

+44 113 244 2829

Teleprinter No.

2. The International Bureau hereby notifies the applicant that the following change has been recorded concerning:

 the person the name the address the nationality the residence

Name and Address

REDDIE & GROSE
Daedalus House
Station Road
Cambridge CB1 2RE
United Kingdom

State of Nationality

State of Residence

Telephone No.

+44 (0) 1223 360350

Facsimile No.

+44 (0) 1223 360280

Teleprinter No.

3. Further observations, if necessary:

4. A copy of this notification has been sent to:

 the receiving Office the designated Offices concerned the International Searching Authority the elected Offices concerned the International Preliminary Examining Authority other:The International Bureau of WIPO
34, chemin des Colombettes
1211 Geneva 20, Switzerland

Authorized officer

Dominique DELMAS

Facsimile No.: (41-22) 740.14.35

Telephone No.: (41-22) 338.83.38

THIS PAGE BLANK (USPTO)

PART II COOPERATION TREATY

From the INTERNATIONAL BUREAU

PCT

NOTIFICATION OF ELECTION
(PCT Rule 61.2)

Date of mailing (day/month/year) 10 November 2000 (10.11.00)	To: Commissioner US Department of Commerce United States Patent and Trademark Office, PCT 2011 South Clark Place Room CP2/5C24 Arlington, VA 22202 ETATS-UNIS D'AMERIQUE in its capacity as elected Office
International application No. PCT/GB00/00582	Applicant's or agent's file reference RCD/P38040WO
International filing date (day/month/year) 18 February 2000 (18.02.00)	Priority date (day/month/year) 20 February 1999 (20.02.99)
Applicant ANDREWS, Peter et al	

1. The designated Office is hereby notified of its election made:

 in the demand filed with the International Preliminary Examining Authority on:

08 September 2000 (08.09.00)

 in a notice effecting later election filed with the International Bureau on:

2. The election was was not

made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland Facsimile No.: (41-22) 740.14.35	Authorized officer Dominique DELMAS Telephone No.: (41-22) 338.83.38
---	--

THIS PAGE BLANK (USPTO)

PATENT COOPERATION TREATY

From the INTERNATIONAL BUREAU

PCT

NOTIFICATION OF THE RECORDING
OF A CHANGE(PCT Rule 92bis.1 and
Administrative Instructions, Section 422)

Date of mailing (day/month/year) 17 October 2000 (17.10.00)	To: REDDIE & GROSE 16, Theobalds Road London WC1X 8PL ROYAUME-UNI											
Applicant's or agent's file reference RCD/P38040WO	IMPORTANT NOTIFICATION											
International application No. PCT/GB00/00582	International filing date (day/month/year) 18 February 2000 (18.02.00)											
<p>1. The following indications appeared on record concerning:</p> <p><input type="checkbox"/> the applicant <input type="checkbox"/> the inventor <input checked="" type="checkbox"/> the agent <input type="checkbox"/> the common representative</p> <table border="1"> <tr> <td rowspan="4">Name and Address HARRISON GODDARD FOOTE Tower House Merrion Way Leeds LS2 8PA United Kingdom</td> <td>State of Nationality</td> <td>State of Residence</td> </tr> <tr> <td colspan="2">Telephone No. +44 1223 360350</td> </tr> <tr> <td colspan="2">Facsimile No. +44 1223 360280</td> </tr> <tr> <td colspan="2">Teleprinter No.</td> </tr> </table>				Name and Address HARRISON GODDARD FOOTE Tower House Merrion Way Leeds LS2 8PA United Kingdom	State of Nationality	State of Residence	Telephone No. +44 1223 360350		Facsimile No. +44 1223 360280		Teleprinter No.	
Name and Address HARRISON GODDARD FOOTE Tower House Merrion Way Leeds LS2 8PA United Kingdom	State of Nationality	State of Residence										
	Telephone No. +44 1223 360350											
	Facsimile No. +44 1223 360280											
	Teleprinter No.											
<p>2. The International Bureau hereby notifies the applicant that the following change has been recorded concerning:</p> <p><input checked="" type="checkbox"/> the person <input type="checkbox"/> the name <input type="checkbox"/> the address <input type="checkbox"/> the nationality <input type="checkbox"/> the residence</p> <table border="1"> <tr> <td rowspan="4">Name and Address REDDIE & GROSE 16, Theobalds Road London WC1X 8PL United Kingdom</td> <td>State of Nationality</td> <td>State of Residence</td> </tr> <tr> <td colspan="2">Telephone No. +44 1223 360350</td> </tr> <tr> <td colspan="2">Facsimile No. +44 1223 360280</td> </tr> <tr> <td colspan="2">Teleprinter No.</td> </tr> </table>				Name and Address REDDIE & GROSE 16, Theobalds Road London WC1X 8PL United Kingdom	State of Nationality	State of Residence	Telephone No. +44 1223 360350		Facsimile No. +44 1223 360280		Teleprinter No.	
Name and Address REDDIE & GROSE 16, Theobalds Road London WC1X 8PL United Kingdom	State of Nationality	State of Residence										
	Telephone No. +44 1223 360350											
	Facsimile No. +44 1223 360280											
	Teleprinter No.											
<p>3. Further observations, if necessary:</p>												
<p>4. A copy of this notification has been sent to:</p> <table> <tr> <td><input checked="" type="checkbox"/> the receiving Office</td> <td><input checked="" type="checkbox"/> the designated Offices concerned</td> </tr> <tr> <td><input checked="" type="checkbox"/> the International Searching Authority</td> <td><input type="checkbox"/> the elected Offices concerned</td> </tr> <tr> <td><input type="checkbox"/> the International Preliminary Examining Authority</td> <td><input checked="" type="checkbox"/> other: Former Agent</td> </tr> </table>				<input checked="" type="checkbox"/> the receiving Office	<input checked="" type="checkbox"/> the designated Offices concerned	<input checked="" type="checkbox"/> the International Searching Authority	<input type="checkbox"/> the elected Offices concerned	<input type="checkbox"/> the International Preliminary Examining Authority	<input checked="" type="checkbox"/> other: Former Agent			
<input checked="" type="checkbox"/> the receiving Office	<input checked="" type="checkbox"/> the designated Offices concerned											
<input checked="" type="checkbox"/> the International Searching Authority	<input type="checkbox"/> the elected Offices concerned											
<input type="checkbox"/> the International Preliminary Examining Authority	<input checked="" type="checkbox"/> other: Former Agent											

The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland Facsimile No.: (41-22) 740.14.35	Authorized officer Eugénia Santos Telephone No.: (41-22) 338.83.38
---	--

THIS PAGE BLANK (USPTO)

PATENT COOPERATION TREATY

PCT

NOTIFICATION OF THE RECORDING
OF A CHANGE(PCT Rule 92bis.1 and
Administrative Instructions, Section 422)Date of mailing (day/month/year)
16 October 2000 (16.10.00)

From the INTERNATIONAL BUREAU

To:

REDDIE & GROSE
16, Theobalds Road
London WC1X 8PL
ROYAUME-UNI

Date of mailing (day/month/year) 16 October 2000 (16.10.00)	
Applicant's or agent's file reference RCD/P38040WO	IMPORTANT NOTIFICATION
International application No. PCT/GB00/00582	International filing date (day/month/year) 18 February 2000 (18.02.00)

1. The following indications appeared on record concerning:

the applicant the inventor the agent the common representative

Name and Address UNIVERSITY OF SHEFFIELD Western Bank Sheffield S10 2TN United Kingdom	State of Nationality GB	State of Residence GB
	Telephone No.	
	Facsimile No.	
	Teleprinter No.	

2. The International Bureau hereby notifies the applicant that the following change has been recorded concerning:

the person the name the address the nationality the residence

Name and Address INTERCYTEX LIMITED Incubator Building Grafton Street Manchester, M13 9XX United Kingdom	State of Nationality GB	State of Residence GB
	Telephone No.	
	Facsimile No.	
	Teleprinter No.	

3. Further observations, if necessary:

4. A copy of this notification has been sent to:
<input checked="" type="checkbox"/> the receiving Office <input checked="" type="checkbox"/> the designated Offices concerned <input checked="" type="checkbox"/> the International Searching Authority <input type="checkbox"/> the elected Offices concerned <input type="checkbox"/> the International Preliminary Examining Authority <input type="checkbox"/> other:

The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland Facsimile No.: (41-22) 740.14.35	Authorized officer Eugénia Santos Telephone No.: (41-22) 338.83.38
---	--

THIS PAGE BLANK (USPTO)

From the
INTERNATIONAL PRELIMINARY EXAMINING AUTHORITY

To:

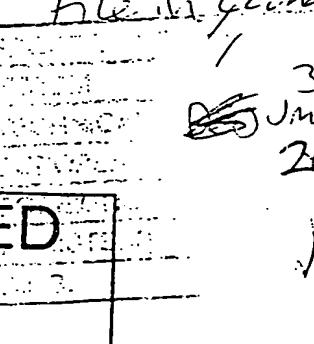
REDDIE & GROSE
16 Theobalds Road
LONDON, WC1X 9PL
GRANDE BRETAGNE

RECEIVED

29 JUN 2001

CAMBRIDGE
RCD/P38046WO

1555



PCT

NOTIFICATION OF TRANSMITTAL OF
THE INTERNATIONAL PRELIMINARY
EXAMINATION REPORT

(PCT Rule 71.1)

Date of mailing
(day/month/year) 05.06.2001

IMPORTANT NOTIFICATION

International application No.
PCT/GB00/00582

International filing date (day/month/year)
18/02/2000

Priority date (day/month/year)
20/02/1999

Applicant
UNIVERSITY OF SHEFFIELD et al.

1. The applicant is hereby notified that this International Preliminary Examining Authority transmits herewith the international preliminary examination report and its annexes, if any, established on the international application.
2. A copy of the report and its annexes, if any, is being transmitted to the International Bureau for communication to all the elected Offices.
3. Where required by any of the elected Offices, the International Bureau will prepare an English translation of the report (but not of any annexes) and will transmit such translation to those Offices.
4. REMINDER

The applicant must enter the national phase before each elected Office by performing certain acts (filing translations and paying national fees) within 30 months from the priority date (or later in some Offices) (Article 39(1)) (see also the reminder sent by the International Bureau with Form PCT/IB/301).

Where a translation of the international application must be furnished to an elected Office, that translation must contain a translation of any annexes to the international preliminary examination report. It is the applicant's responsibility to prepare and furnish such translation directly to each elected Office concerned.

For further details on the applicable time limits and requirements of the elected Offices, see Volume II of the PCT Applicant's Guide.

CAMBRIDGE

DATE 20.8.01

CAN

Name and mailing address of the IPEA/



European Patent Office
D-80298 Munich
Tel. +49 89 2399 - 0 Tx: 523656 epmu d
Fax: +49 89 2399 - 4465

Authorized officer

CLEERE, C.

Tel. +49 89 2399-8061



THIS PAGE BLANK (USPTO)

PATENT COOPERATION TREATY

PCT

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

Applicant's or agent's file reference RCD/P38040WO	FOR FURTHER ACTION		See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)
International application No. PCT/GB00/00582	International filing date (day/month/year) 18/02/2000	Priority date (day/month/year) 20/02/1999	
International Patent Classification (IPC) or national classification and IPC C12N5/22			
Applicant UNIVERSITY OF SHEFFIELD et al.			
<p>1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.</p> <p>2. This REPORT consists of a total of 8 sheets, including this cover sheet.</p> <p><input checked="" type="checkbox"/> This report is also accompanied by ANNEXES, i.e. sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).</p> <p>These annexes consist of a total of 5 sheets.</p>			
<p>3. This report contains indications relating to the following items:</p> <ul style="list-style-type: none"> I <input checked="" type="checkbox"/> Basis of the report II <input checked="" type="checkbox"/> Priority III <input type="checkbox"/> Non-establishment of opinion with regard to novelty, inventive step and industrial applicability IV <input type="checkbox"/> Lack of unity of invention V <input checked="" type="checkbox"/> Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement VI <input type="checkbox"/> Certain documents cited VII <input type="checkbox"/> Certain defects in the international application VIII <input checked="" type="checkbox"/> Certain observations on the international application 			

Date of submission of the demand 09/08/2000	Date of completion of this report 05.06.2001
Name and mailing address of the international preliminary examining authority: European Patent Office D-80298 Munich Tel. +49 89 2399 - 0 Tx: 523656 epmu d Fax: +49 89 2399 - 4465	Authorized officer Mundel, C Telephone No. +49 89 2399 7314



THIS PAGE BLANK (USPTO)

INTERNATIONAL PRELIMINARY
EXAMINATION REPORT

International application No. PCT/GB00/00582

I. Basis of the report

1. With regard to the elements of the international application (*Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to this report since they do not contain amendments (Rules 70.16 and 70.17)*):
Description, pages:

1-44 as originally filed

Claims, No.:

1-27 as received on 19/03/2001 with letter of 15/03/2001

Drawings, sheets:

1/11-11/11 as originally filed

2. With regard to the language, all the elements marked above were available or furnished to this Authority in the language in which the international application was filed, unless otherwise indicated under this item.

These elements were available or furnished to this Authority in the following language: , which is:

- the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
- the language of publication of the international application (under Rule 48.3(b)).
- the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).

3. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international preliminary examination was carried out on the basis of the sequence listing:

- contained in the international application in written form.
- filed together with the international application in computer readable form.
- furnished subsequently to this Authority in written form.
- furnished subsequently to this Authority in computer readable form.
- The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
- The statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished.

4. The amendments have resulted in the cancellation of:

- the description, pages:
- the claims, Nos.:

THIS PAGE BLANK (USPTO)

INTERNATIONAL PRELIMINARY
EXAMINATION REPORT

International application No. PCT/GB00/00582

the drawings, sheets:

5. This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):

(Any replacement sheet containing such amendments must be referred to under item 1 and annexed to this report.)

6. Additional observations, if necessary:

II. Priority

1. This report has been established as if no priority had been claimed due to the failure to furnish within the prescribed time limit the requested:

copy of the earlier application whose priority has been claimed.

translation of the earlier application whose priority has been claimed.

2. This report has been established as if no priority had been claimed due to the fact that the priority claim has been found invalid.

Thus for the purposes of this report, the international filing date indicated above is considered to be the relevant date.

3. Additional observations, if necessary:
see separate sheet

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty (N)	Yes:	Claims 1-19, 21-22 and 24-27 (See Citations and explanations)
	No:	Claims 20 and 23
Inventive step (IS)	Yes:	Claims 1-19, 21-22 and 24-27 (See Citations and explanations)
	No:	Claims 20 and 23
Industrial applicability (IA)	Yes:	Claims 1-25 and 27
	No:	Claims 26 (See Citations and explanations)

2. Citations and explanations
see separate sheet

VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:

THIS PAGE BLANK (USPTO)

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT**

International application No. PCT/GB00/00582

see separate sheet

THIS PAGE BLANK (USPTO)

Re Item II

Priority

The priority document of the present application was not available at the time where this International Preliminary Examination Report (IPER) has been drafted. The present analysis is based on the hypothesis that all the claims have a priority right corresponding to the date of filing of the priority document (20.02.99).

Re Item V

Reasoned statement under Rule 66.2(a)(ii) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. The present application refers to a cell comprising at least part of the cytoplasm derived from an embryonal teratocarcinoma cell combined with a nucleus of a somatic cell, to a cell-line comprising such a cell, a tissue-type or organ comprising such a cell. The application refers to methods for the preparation of a cytoplasmic part of an embryonal teratocarcinoma cell, methods for the preparation of a cell according to the present application, methods for inducing differentiation of a cell according to the present application. Finally, the present application refers to a method to treat conditions or diseases requiring transplantation of tissue and to therapeutic composition or kit comprising at least one cell according to the present application.

2. Reference is made to the following documents :

D1: WANGH LAWRENCE J ET AL: 'Efficient reactivation of Xenopus erythrocyte nuclei in Xenopus egg extracts.' JOURNAL OF CELL SCIENCE, vol. 108, no. 6, 1995, pages 2187-2196.

D2: TOSU M ET AL: 'CLONAL ISOLATION AND CHARACTERIZATION OF MYOBLAST-LIKE RECONSTITUTED CELLS FORMED BY FUSION OF KARYOPLASTS FROM MOUSE TERATOCARCINOMA CELLS WITH RAT MYOBLAST CYTOPLASTS' CELL STRUCTURE AND FUNCTION, vol. 13, no. 3, 1988, pages 249-266.

The document D1 discloses the reactivation of Xenopus erythrocytes nuclei in

THIS PAGE BLANK (USPTO)

INTERNATIONAL PRELIMINARY
EXAMINATION REPORT - SEPARATE SHEET

International application No. PCT/GB00/00582

Xenopus egg extracts.

The document D2 discloses fusion between karyoplasts from mouse teratocarcinoma cells and rat myoblast cytoplasts.

3. New claims 1-27 filed with the letter of 15.03.01 are allowable under articles 19(2) and 34(2)(b) PCT. The arguments filed by the applicant with the letter of 15.03.01 have been taken into account for drafting the present IPER.
4. **Novelty and inventive step; articles 33(2) and 33(3) PCT.**

The subject-matter of present claims 1-19, 21-22 and 24-27 has never been disclosed or suggested in the documents cited in the International Search Report (ISR). Therefore, claims 1-19, 21-22 and 24-27 are considered as new and inventive in the sense of articles 33(2) and 33(3) PCT.

Due to the clarity problems mentioned in points VIII-3 and VIII-5 of the present IPER, the International Preliminary Examination Authority (IPEA) considers that the subject-matter of claim 20 can not be considered as new over well-known pluripotential cell lines and that the subject-matter of claim 23 can not be considered as new over tissues or organs comprising well-known pluripotential cells.

The attention of the applicant is drawn to the fact that the novelty and inventive step of the claims have been determined on the basis that a cell comprising at least part of the cytoplasm derived from an embryonal teratocarcinoma cell combined with the nucleus of a somatic cell will have the pluripotential characteristics disclosed in the present application. However, the IPEA emits doubts about the fact that a cell comprising "part of the cytoplasm of an embryonic teratocarcinoma cell" would acquire the pluripotential characteristic(s) claimed for the cells of the present application (see point VIII-1 beneath).

THIS PAGE BLANK (USPTO)

5. Industrial applicability; article 33(4) PCT.

Claim 26 of the present application is directed to a method of treatment of the human or animal body.

For the assessment of the present claim 26 on the question whether it is industrially applicable, no unified criteria exist in the PCT Contracting States. The patentability can also be dependent upon the formulation of the claims. The EPO, for example, does not recognize as industrially applicable the subject-matter of claims to the use of a compound in medical treatment, but may allow, however, claims to a known compound for first use in medical treatment and the use of such a compound for the manufacture of a medicament for a new medical treatment.

Re Item VIII

Certain observations on the international application

1. As a general remark, the attention of the applicant is drawn to the fact that, since the only examples given in the present application refer to heterokaryons, there is no proof in the present application that a cell comprising a part or even all the cytoplasm from an embryonal teratocarcinoma cell but without the nucleus of said embryonal teratocarcinoma cell combined with a nucleus of a somatic cell could have the "pluripotential characteristic" disclosed in the present application. Therefore, the IPEA considers that all the claims referring to such cells can not be considered as supported by the description of the present application (article 5 PCT in combination with article 6 PCT).
2. In claim 1, the use of the term "derived" renders the scope of the claim since nothing is said about the nature of the "derivation".
3. Claim 20 refers to "a cell culture comprising at least one cell according to the invention". The IPEA considers that the scope of that claim is unclear because no means to distinguish a cell according to the present application from another pluripotential cell, like an ES cell for example, are given. Therefore, the skilled person will not be able to distinguish a cell culture according to claim 20 from an ES cell culture for example.

THIS PAGE BLANK (USPTO)

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT - SEPARATE SHEET**

International application No. PCT/GB00/00582

4. Claim 21 of the present application refers to a method for inducing differentiation of at least one cell according to claims 1-12. The attention of the applicant is drawn to the fact that there are no proof in the present application that a cell according to the present application, even the heterokaryon, could be induced to differentiate in a specific tissue (see also point VIII-1 above). Thus, the IPEA considers that claim 21 is not supported by the description of the present application (article 5 PCT in combination with article 6 PCT). This remark also applies to claim 22.
5. Claim 23 refers to at least one tissue type or organ comprising at least one cell according to any of claims 1-10. Due to the clarity problem mentioned in point VIII-3 above, the IPEA is the opinion that it would not be possible to distinguish an organ or tissue type according to claim 23 from organs or tissues comprising other pluripotential cells. Moreover, the attention of the applicant is drawn to the fact that during the Regional Phase before the EPO, an objection under Rule 23e EPC could be raised against present claim 23.
6. Claim 25 refers to a therapeutical composition comprising at least one cell according to any of claims 1-12 for use in tissue transplantation. The attention of the applicant is drawn to the following facts :
 - (i) A claim to a substance or composition for a particular use is construed as meaning a substance or composition which is in fact suitable for the stated use; a known product which is per se the same as the substance or composition defined in the claim, but which is in a form which would render it unsuitable for the stated use, would not deprive the claim of novelty (PCT International Preliminary Examination Guidelines, as in force from 09.10.98, Section IV, paragraph III-4.8).
 - (ii) There is no proof in the present application that a cell according to claims 1-10 could be used in tissue transplantation. Therefore, the IPEA considers that claim 25 is not supported by the description of the present application (article 5 PCT in combination with article 6 PCT). This remark is also valid for claim 26.

THIS PAGE BLANK (USPTO)

- 41 -

CLAIMS

1. A cell having a single nucleus, which cell possesses at least one pluripotential characteristic, which characteristic includes the ability to differentiate into one of at least two selected tissue types, which cell comprises either (i) at least part of the cytoplasm derived from a embryonal teratocarcinoma cell, or (ii) a cytoplasm from a embryonal teratocarcinoma cell, and which cell has its nucleus obtained from a differentiated somatic cell.
2. A cell according to Claim 1 characterised in that said pluripotential characteristic includes the ability of said cell to proliferate in culture in an undifferentiated state.
3. A cell according to Claim 2 characterised in that said cell has the capacity to proliferate in continuous culture in an undifferentiated state for at least six months and ideally 12 months.
4. A cell according to any of Claims 1-3 characterised in that said pluripotential characteristic includes the expression of at least one selected marker.
5. A cell according to Claim 4 characterised in that said pluripotential characteristic is expression of Oct4.
6. A cell according to Claim 4 characterised in that said selected marker is a cell surface marker.
7. A cell according to Claim 6 characterised in that

THIS PAGE BLANK (USPTO)

- 42 -

said cell surface marker is selected from the group including; SSEA-1 (-); and/or SSEA-3 (+); and/or SSEA-4 (+); and/or TRA-1-60 (+); and/or TRA-1-81 (+); and/or alkaline phosphatase (+).

5 8. A cell according to Claims 1-7 characterised in that said pluripotential characteristic includes the presence of telomerase activity.

9. A cell according to any of Claims 1-8 characterised in that said pluripotential characteristic includes the 10 presence of a chromosomal methylation pattern characteristic of pluripotential cells.

10. A cell according to any of Claim 1-9 characterised in that said pluripotential characteristic includes the ability to induce tumours when introduced into an animal.

15 11. A cell-line consisting of cells according to any of Claims 1-10.

12. A cell-line according to Claim 11 characterised in that said cell-line is of human origin.

13. A method for the preparation of a cytoplasmic part for 20 use in the production of a cell according to any of Claims 1-10 or a cell-line according to Claims 11 or 12 comprising;

- (i) providing at least one embryonal teratocarcinoma cell;
- (ii) separating at least part of the cytoplasm from the nucleus of said cell;

THIS PAGE BLANK (USPTO)

- 43 -

- (iii) isolating said cytoplasmic part; and,
optionally
- (iv) storing said isolated cytoplasmic part under
suitable storage conditions.

14. A method according to Claim 13 characterised in that
said cytoplasmic part is a cytoplasm.

15. A method for preparing a cell according to any of
Claims 1-10 or a cell-line according to Claims 11 or 12
comprising;

- (i) combining at least one embryonal
teratocarcinoma cell with at least one
differentiated somatic cell;
- (ii) removing the embryonal teratocarcinoma nucleus
from said combined cell;
- (iii) culturing said cell under conditions conducive
to proliferation and expansion of said cell;
and, optionally
- (iv) storing said cell culture under suitable
conditions.

16. A method for combining at least part of the cytoplasm
of an embryonal teratocarcinoma cell with a somatic cell
comprising;

- (i) providing at least part of the cytoplasm of an
embryonal teratocarcinoma cell;
- (ii) combining said cytoplasmic part with at least
one differentiated somatic cell;
- (iii) growing said combined cell in culture; and
optionally

THIS PAGE BLANK (USPTO)

- 44 -

(iv) storing said combined cell under suitable storage conditions.

17. A method according to Claim 16 characterised in that said cytoplasmic part is provided as a cytoplasm.

5 18. A method according to Claim 16 or 17 characterised in that said cytoplasm is combined with said somatic cell via cytoplasm/somatic cell fusion.

10 19. A method according to Claims 16-18 characterised in that said embryonal carcinoma cell and somatic cell are of human origin.

20. A cell culture comprising at least one cell according to any of Claims 1-10.

15 21. A method for inducing differentiation of at least one cell according to any of Claims 1-10 comprising;

- (i) providing a cell according to any of Claims 1-10;
- (ii) culturing said cell under conditions conducive to the differentiation of said cell into at least one tissue; and optionally
- (iii) storing of said differentiated tissue prior to use under suitable storage conditions.

25 22. A method of Claim 21 characterised in that said method provides a tissue type selected from at least one of the following; neural, smooth muscle, striated muscle, cardiac muscle, bone, cartilage, liver, kidney,

THIS PAGE BLANK (USPTO)

- 45 -

respiratory epithelium haematopoietic cells, spleen, skin, stomach, intestine.

23. At least one tissue type or organ comprising at least one cell according to any of Claims 1-10.

5 24. A therapeutic composition comprising at least one cell according to any of Claims 1-10 including a suitable excipient, diluent or carrier.

25. A therapeutic composition according to Claim 24 characterised in that said therapeutic composition is 10 provided for use in tissue transplantation.

26. A method to treat conditions or diseases requiring transplantation of tissue comprising:

(i) providing at least one tissue type or organ according to Claim 23;

15 (ii) surgically introducing said tissue or organ into a patient to be treated; and

(iii) treating said patient under conditions which are conducive to the acceptance of said transplanted tissue by said patient.

20 27. A kit comprising at least one cell according to any of Claims 1-10; instructions with respect to maintenance of said cell in culture; and, optionally, factors required to induce differentiation of said cell to at least one desired tissue type or organ.

THIS PAGE BLANK (USPTO)

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau



41555

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁷ : C12N 5/22, A61K 35/12, 35/54, A61P 43/00 // C12N 5/28		A2	(11) International Publication Number: WO 00/49138 (43) International Publication Date: 24 August 2000 (24.08.00)
(21) International Application Number: PCT/GB00/00582		(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).	
(22) International Filing Date: 18 February 2000 (18.02.00)		(30) Priority Data: 9903805.1 20 February 1999 (20.02.99) GB	
(71) Applicant (for all designated States except US): UNIVERSITY OF SHEFFIELD [GB/GB]; Western Bank, Sheffield S10 2TN (GB).		(72) Inventors; and (75) Inventors/Applicants (for US only): ANDREWS, Peter [GB/GB]; University of Sheffield, Dept. of Biomedical Sciences, Wester Bank, Sheffield S10 2TN (GB). KEMP, Paul [GB/GB]; 16 Chadkirk Road, Romiley SK6 3JY (GB).	
(74) Agent: HARRISON GODDARD FOOTE; Tower House, Merrion Way, Leeds LS2 8PA (GB).		Published <i>Without international search report and to be republished upon receipt of that report.</i>	

(54) Title: PLURIPOTENTIAL CELLS-1

(57) Abstract

The invention relates to isolated pluripotential cells comprising at least part of the cytoplasm from a teratocarcinoma cell and a nucleus of a somatic cell. The invention also relates to methods to prepare such cells.

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
AU	Australia	GA	Gabon	LV	Latvia	SZ	Swaziland
AZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav Republic of Macedonia	TM	Turkmenistan
BF	Burkina Faso	GR	Greece	ML	Mali	TR	Turkey
BG	Bulgaria	HU	Hungary	MN	Mongolia	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MR	Mauritania	UA	Ukraine
BR	Brazil	IL	Israel	MW	Malawi	UG	Uganda
BY	Belarus	IS	Iceland	MX	Mexico	US	United States of America
CA	Canada	IT	Italy	NE	Niger	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NL	Netherlands	VN	Viet Nam
CG	Congo	KE	Kenya	NO	Norway	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NZ	New Zealand	ZW	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's Republic of Korea	PL	Poland		
CM	Cameroon	KR	Republic of Korea	PT	Portugal		
CN	China	KZ	Kazakhstan	RO	Romania		
CU	Cuba	LC	Saint Lucia	RU	Russian Federation		
CZ	Czech Republic	LI	Liechtenstein	SD	Sudan		
DE	Germany	LK	Sri Lanka	SE	Sweden		
DK	Denmark	LR	Liberia	SG	Singapore		
EE	Estonia						

518 Rec'd PCT/PTO 20 AUG 2001

7/prs

PLURIPOTENTIAL CELLS-1

The invention herein described relates to isolated pluripotential cells, 5 comprising at least part of the cytoplasm from a teratocarcinoma cell and a nucleus of a somatic cell; methods to prepare such cells; therapeutic compositions of said cells; and uses thereof.

Animal embryonic development is a highly regulated development process 10 that combines cell proliferation and cell/tissue differentiation to produce an intact organism. The co-ordination of cell proliferation and differentiation is, and has been, the subject of intense research and the information derived from this has contributed to our understanding of cell function and disease. For example and not by way of limitation, regulation of gene expression, cell 15 differentiation, oncology, teratology.

Mammalian embryonic development is remarkably conserved during the early stages. Post fertilisation the early embryo completes four rounds of cleavage to form a morula of 16 cells. These cells complete several more rounds of 20 division and develop into a blastocyst in which the cells can be divided into two distinct regions; the inner cell mass, which will form the embryo, and the trophectoderm, which will form extra embryonic tissue, (eg placenta).

Those cells that form part of the embryo up until the formation of the 25 blastocyst are said to be totipotent (e.g. each cell has the developmental potential to form a complete embryo and all the cells required to support the growth and development of said embryo).

During the formation of the blastocyst, the cells that comprise the inner cell mass (ICM) are said to be pluripotential (ie each cell has the developmental potential to form a variety of tissues).

- 5 Embryonic stem cells may be principally derived from two embryonic sources. Pluripotential cells isolated from the inner cell mass are termed embryonic stem cells (ES cells). An alternate source of pluripotential cells is derived from primordial germ cells isolated from the mesenteries or genital ridges of days 8.5-12.5 *post coitum* embryos which would ultimately 10 differentiate into germ cells. These pluripotential cells are referred to as embryonic germ cells (EG cells). Each of these types of pluripotential cell has the similar developmental potential with respect to differentiation into alternate cell types.
- 15 It is important to note that an intact embryo cannot be produced from a single pluripotential cell (eg either an ES or EG cell). Therefore a pluripotential cell has an increased commitment to terminal differentiation when compared to a totipotent cell.
- 20 Until very recently *in vitro* culture of human ES cells was not possible. The first indication that conditions may be determined which could allow the establishment of human ES cells in culture is described in WO 96/22362. The application describes cell lines and growth conditions which allow the continuous proliferation of primate ES cells which exhibit a range of 25 characteristics or markers which are associated with stem cells having pluripotent characteristics.

For example, and not by way of limitation, the expression of specific cell surface markers SSEA-3 (+), SSEA-4 (+), TRA-1-60 (+), TRA-1-81 (+) (

- 30 Shevinsky *et al* 1982; Kannagi *et al* 1983; Andrews *et al* 1984a; Thomson *et*

al 1995) and alkaline phosphatase (+). In addition the established primate cell lines disclosed in WO 96/22362 have stable karyotypes and continue to proliferate in an undifferentiated state in continuous culture. The primate ES 5 cell lines also retain the ability, throughout their continuous culture, to form tissues derived from all three embryonic germ layers (endoderm, mesoderm and ectoderm).

More recently Thomson *et al* (Science 282: 1145-1147, 1998) have published 10 conditions in which human ES cells can be established in culture. The above characteristics shown by primate ES cells are also shown by the human ES cell lines. In addition the human cell lines show high levels of telomerase activity, a characteristic of cells which show the ability to divide continuously in culture.

15

An alternative source of pluripotential embryonic stem cells are those derived from primordial embryonal germ cells (EG cells) which are located in the mesenteries or genital ridges of embryos. WO 98/43679 describes the isolation 20 of EG cells from the gonadal or genital ridges of human embryos. EG cells described in WO 98/43679 exhibit features in common with primate and human ES cells, (eg expression of cell surface markers, continuous proliferation in culture in an undifferentiated state, normal karyotype and the ability to differentiate into selected tissues under defined conditions).

25 It is evident that the use of *in vitro* cultures of pluripotential stem cells, especially human cells, has important ramifications for both basic research (eg as a model for studying gene expression and/or tissue differentiation) and in transplantation and/or replacement therapies for tissues which have been damaged either through injury or disease. The establishment of *in vitro* 30 cultures of human ES and EG cells is a major step toward realising the full

potential of this technology; because of their pluripotent nature ES and EG cells may be capable of differentiating under controlled conditions into a variety of cell types and/or tissues and organs that could have a wide variety of applications. For example, and not by way of limitation, replacement of 5 damaged and/or diseased coronary and/or major arteries; replacement of damaged and/or diseased organs (eg as a result of kidney disease, (eg cirrhosis), diabetes, various autoimmune diseases); replacement of damaged neurones (eg Alzhiemers disease, Parkinsons disease, spinal injuries) or cancer. It will also be apparent to one skilled in the art that diseases such as 10 AIDS may benefit from tissues derived from ES or EG cells. The depletion of T-cells through virus induced cell death is the major contributory factor to the immuno-compromised state of AIDS sufferers.

However, there are practical and ethical difficulties associated with the use of 15 material derived from human embryos. Moreover, such allogeneic material, if transplanted into another human, may illicit a severe immune reaction in the host and be thus destroyed.

It has been known for many years that amphibian somatic cell nuclei retain 20 their ability to give rise to entire organisms when they are transplanted into egg cells which have had their nucleus removed or inactivated (Gurdon 1974). Thus determination of the pluripotent of these cells must be controlled by the egg cytoplasm which was able to in effect reprogramme the somatic cell nucleus into a totipotent state.

25

Mammalian somatic cell nuclei have also been shown to retain this placidity and can be reprogrammed when transferred to enucleated oocytes, (Campbell *et al*; Wakayama *et al*)

Moreover nucleated mouse ES cells have been shown to be able to reprogramme somatic cell nuclei, although in this case, a heterokaryon was produced containing the cytoplasm and nuclei from both types of cells so it is difficult to determine the actual mechanism of action of the reprogramming state.

In all these examples, although the material produced is genetically identical to the somatic cell donor, these somatic cells were reprogrammed by cellular elements derived from either oocytes or ES cells and again, in human this poses practical and ethical concerns.

Embryonal carcinoma cells derived from teratomas are also able to reprogramme somatic cell nuclei in order to produce cells with pluropotential characteristics.

15

Teratomas, tumours that contain a wide range of more or less organised tissues have been known in humans for many hundreds of years. They typically occur as gonadal tumours of both men and women, but they also occur in other sites. The gonadal forms of these tumours are generally believed to originate from germ cells, and the extra-gonadal forms, which typically have the same range of histology, are widely thought to arise from 'mis-placed' germ cells that have migrated incorrectly during embryogenesis. However, a non-germ cell origin from persisting embryonic stem cells may be considered in some cases, especially for teratomas occurring in the new born.

25

Because of the presumption of a germ cell origin in most cases, teratomas are generally classed as germ cell tumours (GCT), which also manifest a range of other histological types, including seminoma (often called dysgerminoma in females), embryonal carcinoma (EC), yolk sac carcinoma and

choriocarcinoma. GCT may contain any combination of these tissue types, with or without elements of teratoma. Combinations of teratoma and EC are frequently described as teratocarcinoma. Commonly, human GCT are divided into pure seminomas, in which none of the other histological types occur, and 5 non-seminomatous GCT (NSGCT), which may contain any combination of histological types, with or without elements of teratoma. Confusingly, NSGCT may also contain elements of seminoma.

Ovarian GCT are most commonly benign and contain only well differentiated 10 somatic tissues that may include bone, muscle, nerve. Often well-organised tissues are found, including teeth and hair. By contrast, human testicular GCT are always malignant containing any combination of the tumour tissue types discussed above. Any teratoma elements present may be less well organised than in benign human ovarian teratomas, but somatic cell types such as nerve, 15 muscle, bone and cartilage may be quite recognisable. Testicular GCT are rare but have a peak incidence after puberty, being the most common form of cancer in young men between the ages of 20 - 35.

Early histopathological studies of human GCT led to the proposal that EC 20 cells resemble early embryonic cells and are the stem cells that give rise to all the other cell types in GCT, with the exception of seminoma. (In the currently prevailing view, seminoma more closely resembles the primordial germ cells from which these tumours arise, and so may represent an earlier stage in tumour development and progression). Detailed experimental study of GCT 25 became possible with the discovery that male laboratory mice of the strain 129 develop spontaneous testicular teratocarcinomas. Studies of these mice confirmed that these tumours had a germ cell tumour origin, and indeed arose from primordial germ cells (PGC) at about the time that they migrated into the genital ridge of the developing embryo - 11 - 13 days of gestation in the

laboratory mouse. Evidence was also obtained supporting the hypothesis that EC cells are pluripotent stem cells that are able to generate the whole range of differentiated cells found in teratocarcinomas. Subsequently it was found that similar teratocarcinomas and teratomas could be derived from embryos that 5 have been transplanted to ectopic sites.

For reasons that remain unclear, seminoma has not been observed in laboratory mice, and so no animal model of this tumour exists. Spermatocytic seminoma do occur in dogs but, although these also occur in old men, they 10 appear to be a quite different tumour type with different aetiology to the GCT.

EC cell lines that can be maintained in vitro have been derived from several mouse teratocarcinomas. Some of these EC cell lines were found to retain a pluripotent phenotype and could be induced to differentiate into a range of cell 15 types, either by transplantation back to a syngeneic mouse, in which case a teratocarcinoma would be formed, or by various manipulations in vitro. Some EC cell lines differentiated spontaneously when allowed to grow to confluence. Others required a feeder layer of irradiated or mitomycin-treated cells if they were to retain an EC phenotype, and they differentiated 20 spontaneously if removed from the feeder cells. In a number of cases, it was found that maintaining EC cells in suspension culture for several days forced them to form floating clumps of cells that became vesiculated and began to differentiate. These floating clumps were known as embryoid bodies; a wide 25 range of differentiated cell types, including nerve and muscle, would grow out from these if they were subsequently allowed to attach to a tissue culture surface. It was also found that a number of chemical agents, most strikingly retinoic acid, also induced the differentiation of many murine EC cells into a range of cell types. The precise cell types formed in response to any of these treatments depended upon the particular EC cell line.

Access to EC cell lines allowed detailed characterisation of their properties and pattern of marker expression. Several surface antigens, notably the 'F9 antigen' and Stage Specific Embryonic Antigen-1 (SSEA-1) were identified as characteristically expressed by mouse EC cells. It was further noted that these 5 cells did not express class 1 major histocompatibility (MHC) antigens - H-2, in the mouse. These features, as well as morphology and their capacity to differentiate suggested that murine EC cells resembled cells of the inner cell mass or primitive ectoderm of the early mouse embryo. This resemblance was confirmed by the finding that some EC cells could differentiate and participate 10 in normal embryonic development, when transferred to a blastocyst of an early mouse embryo, which was then re-implanted in a pseudo-pregnant mother and allowed to develop to term. In some cases the chimeras developing from such EC cell ↔ embryo combinations were normal with extensive contributions from the EC cell component; in some cases the chimeras subsequently 15 developed teratocarcinomas, indicating that the tumour phenotype of the EC cells had not been fully suppressed. Only in a very small number of cases was germ cell chimerism reported.

EC cells have been shown to have many of the features which characterise ES 20 and/or EG cells. EC cells are relatively easy to establish in culture, express cell surface markers associated with ES and/or EG cells, can be maintained in continuous culture in an undifferentiated state and have the potential to differentiate into selected tissues both *in vivo* and *in vitro*. However what is also evident is that EC cells contain a mutation, or mutations, in genes 25 (oncogenes) which result in the additional undesirable feature that the cells retain the potential to form tumours.

It has been known for several years that selected chemical treatments of cells in culture can result in cells extruding nuclei resulting in the formation of

separate nuclear and cytoplasmic parts termed karyoplasts and cytoplasts, respectfully. It is well known in the art that the separated parts of the cell may be reconstituted via cell fusion. For example, and not by way of limitation it is possible to produce a cytoplasm from one cell and fuse the cytoplasm to a 5 selected cell to form a cytoplasmic hybrid, or as is commonly known, a cybrid. In addition it is also possible to fuse the karyoplast to a selected cell to form a nuclear hybrid. The nuclei fuse after nuclear membrane breakdown during mitosis and reconstitute after cytokinesis to form a polyploid or aneuploid nucleus. These techniques are well known in the art and will not be detailed 10 extensively at this stage.

We have prepared cytoplasts derived from EC cells and fused the cytoplasts to form cybrids with selected somatic cells. The aim of this approach is to re-programme the differentiated somatic cell nucleus, through contact with 15 factors located in the EC cytoplasm, so that, the cybrid de-differentiates and so takes on the characteristic features of a pluripotential cell. This then provides the basis for the establishment of pluripotential cell lines which, upon exposure to various differentiation factors, can lead to the production of selected differentiated tissue for use in, amongst other things, transplantation 20 therapy.

Advantageously there is no requirement to use harvested embryonic cells to derive the cytoplasts. Therefore any ethical issues with respect to the use of embryos in this way is circumvented. In addition the establishment of EC 25 cells in culture is a relatively amenable task when compared to the problems of establishing human stem cell cultures *in vitro*. Finally the removal of the EC cell nucleus from the donating EC cell results in no transfer of genetic material carrying potential oncogenes to the cybrid cell so formed.

10

It is therefore an object of the invention to provide a pluripotential cell that is not derived from embryonic tissue from a primary source.

It is a further object of the invention to provide methods of combining at least 5 part of the cytoplasm of an EC cell with a somatic nucleus.

It is yet a further object of the invention to provide a pluripotential cell having the capacity to differentiate into selected tissues upon exposure to appropriate factors.

10

According to a first aspect of the invention there is provided a cell comprising at least part of the cytoplasm derived from at least one embryonal carcinoma cell combined with at least the nucleus of at least one somatic cell.

15 In a preferred embodiment of the invention said cell, ideally a cybrid, is characterised by the possession of at least one pluripotential characteristic.

We believe that the acquisition of this pluripotential characteristic is as a result of the re-programming of said somatic nucleus.

20

It will be apparent to those skilled in the art that the cell of the invention may be derived, most preferably, by the creation of a cybrid; but an alternative option involves the fusion of a somatic cell with an EC cell. Clearly this latter option is not preferred because of the potentially oncogenic genome (gene) of 25 the donating EC cell. Preferably the further step of removing the EC nucleus is described hereinafter.

Ideally said pluripotential characteristic includes the ability to differentiate into at least one selected tissue type, preferably upon exposure to at least one 30 differentiation factor.

Alternatively, or additionally, said pluripotential characteristic includes the ability of said cell to proliferate in culture in an undifferentiated state.

- 5 In yet a further preferred embodiment of the invention said cell has the capacity to proliferate in continuous culture in an undifferentiated state for at least 6 months and ideally 12 months.

- 10 Alternatively or additionally said pluripotential characteristic includes the expression of at least one selected marker of pluripotential cells.

It is well known in the art that pluripotential cells express a number of genes not typically expressed by differentiated cells. These are valuable tools to monitor whether the EC cytoplasm has re-programmed a somatic cell nucleus.

- 15 One such example is Oct4.

In a preferred embodiment of the invention said selected marker is expression of the Oct4 gene.

- 20 In yet still a further preferred embodiment of the invention said selected marker is a cell surface marker. Preferably said cell surface marker is selected from the group including SSEA-1 (-); SSEA-3 (+); SSEA-4 (+); TRA-1-60 (+); TRA-1-81 (+); alkaline phosphatase (+).

- 25 Alternatively, or additionally, said pluripotential characteristic includes the presence of telomerase activity in said pluripotential cell. Ideally said telomerase activity is correlated with extension of telomeres.

- 30 For the sake of clarity, telomerase enzymes add, *de novo*, repetitive DNA sequences to the ends of chromosomes. These ends are referred to as

12

telomeres. For example the telomeres of human chromosomes contain the sequence '5' TTAGGG 3' repeated approximately 1000 times at their ends. In young, dividing cells the telomeres are relatively long. In aging, or non-dividing cells, the telomeres become shortened and there is a strong 5 correlation between telomere shortening and capacity to proliferate. Methods to increase the length of telomeres to increase proliferative capacity are known in the art and are described in WO9513383.

10 Alternatively, or additionally, said pluripotential characteristic includes the presence of a chromosomal methylation pattern characteristic of pluripotential cells.

15 It is well known in the art that the genome of eukaryotic organisms is variably methylated through the addition of methyl (-CH₃) groups attached to cytosine residues in DNA to form 5'methylcytosine (5'-mC). Methylation is correlated with the control of gene expression. Typically genes that are hypomethylated tend to be highly expressed. Hypermethylation is correlated with reduced gene expression. It will be apparent to one skilled in the art that pluripotential cells 20 will have a typical methylation pattern. This pattern may be analysed at a genomic level or at the level of a specific gene. Methods to analyse the extent of methylation are well known in the art and include, by example and not by way of limitation, restriction enzyme digestion of DNA with methylation sensitive restriction endonucleases followed by Southern blotting and probing with suitable gene probes (Umezawa *et al* 1997).

25

Alternatively or additionally said pluripotential characteristic includes the ability to induce tumours when introduced into an animal, ideally a rodent experimental model. More ideally still said animal is immunosuppressed.

13

According to a second aspect of the invention there is provided a cell-line comprising cells according to the invention. Ideally, said cell-line are of human origin.

5 According to a third aspect of the invention there is provided a method for preparing a cytoplasm, or part thereof, for use in the production of the cell or cell line of invention comprising;

10 i) providing at least one EC cell;

ii) separating at least part of the cytoplasm from the nucleus of said EC cell;

iii) isolating said cytoplasmic part; and, optionally

iv) storing said isolated cytoplasmic part prior to use.

15 In a preferred method of the invention said cytoplasmic part is a cytoplasm.

It will be apparent to one skilled in the art that said cytoplasm may be provided either as an aliquot isolated from at least one EC cell (eg an aliquot extracted from an intact EC cell via micromanipulation techniques) or alternatively, and 20 preferably, said cytoplasmic part may be provided as an isolated cytoplasm.

In a preferred method of the invention said cytoplasm part is separated from said nucleus by exposure to a pharmacologically effective amount of at least one cytochalasin. Ideally cytochalasin B.

25

It is well known in the art that cytochalasin B is an example of a chemical effective at separating the nucleus of a cell from the cytoplasm to form a karyoplast and cytoplasm respectively, (Methods in Enzymology Vol 151, p221-237 1987).

30

According to a fourth aspect of the invention there is provided a method for preparing a cell or cell-line in accordance with the invention comprising;

- i) combining at least one EC cell with at least one somatic cell;
- ii) removing from said combined cell, the EC cell nucleus;
- 5 iii) culturing said cell under conditions conducive to proliferation and expansion of said cell; and, optionally
- iv) storing said cell culture under suitable conditions.

It will be apparent to one skilled in the art that methods of micromanipulation 10 exist that facilitate the removal of nuclei from selected cells. It will be apparent that this method of the invention advantageously provides that ;

- i) the factors produced by the EC cell are continually produced thereby maintaining a steady-state level of factors necessary to reprogramme 15 the somatic cell nucleus; and
- ii) the EC cell nucleus is removed from the combined cell prior to mitosis ensuring oncogenic genes are not transferred.

It will be apparent to those skilled in the art that the nature of the somatic cell 20 selected is not critical to the operation of the invention although the cell-type will be selected so as to optimise or maximise success in terms of production of a cell or cell-line of the invention.

According to a fifth aspect of the invention there is provided a method of 25 combining at least part of the cytoplasm of an EC cell with a somatic cell comprising;

- i) providing at least part of the cytoplasm of an EC cell;
- ii) combining said cytoplasmic part with at least one somatic cell;
- iii) growing said combined cell in culture; and, optionally
- 30 iv) storing said combined cell under suitable storage conditions.

In a preferred method of the invention said cytoplasmic part is provided as a cytoplasm.

5 In yet a further preferred method of the invention said cytoplasm is combined with said somatic cell via cytoplasm/somatic cell fusion.

In the above described methods the EC cell and somatic cell are, ideally of human origin.

10

According to a sixth aspect of the invention there is provided a cell culture comprising at least one cell according to the invention.

According to a seventh aspect of the invention there is provided a method for
15 inducing differentiation of at least one cell of the invention comprising:

- i) providing a cell according to the invention;
- ii) culturing said cell under conditions conducive to the differentiation of said cell into at least one tissue; and, optionally
- 20 iii) storage of said differentiated tissue prior to use under suitable storage conditions.

Ideally said culture conditions are selected so as to provide a tissue type, by example and not by way of limitation, that is, neuronal, muscle (eg smooth, striated, cardiac), bone, cartilage, liver, kidney, respiratory epithelium, 25 haematopoietic cells, spleen, skin, stomach, intestine.

According to a eighth aspect of the invention there is provided at least one tissue type or organ comprising at least one cell according to the invention.

30

It will be apparent to one skilled in the art that differentiated tissue according to the invention may have extensive application with respect to transplantation therapy. For example, and not by way of limitation, replacement of damaged and/or diseased coronary and/or major arteries; replacement of damaged and/or diseased organs (eg as a result of kidney disease, (eg cirrhosis), diabetes, various autoimmune diseases); replacement of damaged neurones (eg Alzhiemers disease, Parkinsons disease, spinal injuries) or cancer. It will also be apparent to one skilled in the art that diseases such as AIDS may benefit from from tissues derived from the cells of the invention. The depletion of T-cells through virus induced cell death is the major contributory factor to the immuno-compromised state of AIDS sufferers. The provision of a non-exhaustive supply of T-cells derived from a non-infected somatic cell from the patient has obvious benefits. Moreover, tissue rejection due to host cell immune responses are likely to be negligible since the somatic nucleus used in the cybrid would ideally be derived from the patient requiring the replacement tissue or organ.

According to an nineth aspect of the invention there is provided a therapeutic composition comprising at least one cell of the invention including a suitable excipient, diluant or carrier.

In a preferred embodiment of the invention said therapeutic composition is provided for use in tissue transplantation.

25 According to a tenth aspect of the invention there is provided a method to treat conditions or diseases requiring transplantation of tissue comprising;

- i) providing at least one tissue type or organ according to the invention;
- ii) surgically introducing said tissue type or organ to a patient to be treated; and

iii) treating said patient under conditions which are conducive to the acceptance of said transplanted tissue by said patient.

According to a eleventh aspect of the invention there is provided a kit 5 comprising; at least one cell according to the invention; instructions with respect to the maintenance of said cell in culture; and, optionally, factors required to induce differentiation of said cell to at least one desired tissue type or organ.

10 An embodiment of the invention will now be described by example only and with reference to the following tables and figures wherein;

Table 1 represents a summary of the human EC cell lines and some of the characteristic features of said human EC cell-lines ;

15

Table 2 represents a summary of the murine EC cell lines used;

Figure 1 shows various human teratocarcinoma-derived cell lines. The characteristic embryonal carcinoma (EC) morphology of several human EC 20 cell lines derived from testicular (a-e) and extragonadal (f-g) teratocarcinomas: (a) 1156QE, (b) TERA-1, (c) SuSa, (d) 833KE, (e) 1777NRpmet, (f) 1618K, (g) NCCIT. Bar = 50 μ m;

Figure 2 shows a flow cytometric analyses of surface antigen expression 25 by the human EC cell line 2102Ep;

Figure 3 shows differentiation of several human EC cell lines caused by alteration in growth conditions: (a) 2102Ep cells placed at 10^5 per 75 cm² flask (low density); (b) spontaneous differentiation of a few cells in a culture of 30 1156QE; (c) a culture of SuSa cells passaged by trypsinisation; (d) a culture of

1777NRp differentiated cells derived from 1777NRpmet EC cells by repeated passage of low density (Bronson et al 1983a). Bar =50 μ m; and

Figure 4 shows differentiation of TERA-2 derived human EC cells: (a) 5 NTERA-2 cl D1 human EC cells; (b) parental TERA-2 culture, showing mixed patches of EC and non-EC cells; (c) neurons and other differentiated cells arising in cultures of NTERA-2 cl D1 cells following induction with retinoic acid (Andrews 1984); (d) non-neural differentiated NTERA-2 cl D1 cells induced by exposure to hexamethylene bisacetamide. (Bar =50 μ m);

10

Figure 5 shows heterokaryons obtained by fusion of 2102Ep cl 4D3 human EC cells and the human T-cell leukaemia cell line, MOLT4. Several heterokaryons containing 2 and 3 nuclei can be seen in this field. (Bar = 50 μ m);

15 Figure 6 shows cytoplasts derived from NTERA-2 cl D1 human EC cells following enucleation with cytochalasin B. A remaining nucleated cell (top right) has been included in the field for comparison; and

Figure 7 shows PCR amplification of Oct4 mRNA from a human EC x somatic cell (thymocyte) heterokaryon.

20

Materials and Methods

Preparation of Mouse Thymocytes

The thymocytes were obtained by mincing a thymus removed from a 4-6 week 25 old male mouse (Swiss strain) and suspending the released cells in 10 ml medium (DMEM) with 10% foetal calf serum (FCS). After standing for 2-3 minutes to allow large fragments of thymus to settle, the supernatant was removed and centrifuged at 1500 rpm for 5 min to pellet the suspended

19

thymocytes. The thymocytes were resuspended in fresh medium without FCS, and pelleted again by centrifugation; this was repeated a second time after which the cells were resuspended in fresh serum free medium and counted. Human EC cells were obtained by trypsinisation of confluent cultures as 5 previously described (Andrews *et al.*, 1980; 1982). After washing two times in serum free DMEM, and counting, the human EC cells were mixed with the mouse thymocytes in a ratio of 1 EC cell to 10 thymocytes. The mixed cells were pelleted by centrifugation at 1500 rpm for 5 min.

10

15 Heterokaryon Fusion of Human EC cells and Mouse Thymocytes & Extraction of RNA

The cells were fused using polyethylene glycol (PEG) (Kennett, 1979). The pellet (in Experiment 1, 2×10^6 EC cells and 2×10^7 thymocytes; in Experiment 2, 3×10^6 EC cells and 3×10^7 thymocytes) was resuspended in 20 $200 \mu\text{l}$ 50% (w/v) PEG 1500 in 75 mM HEPES, pH8.0 (Boehringer Mannheim) and incubated at 37° C for 1.5 min. Serum free medium, pre-warmed to 37° C, was then added gradually over 5 min. The cells were then pelleted by centrifugation at 1500 rpm for 5 min. and resuspended in 5 ml DMEM with 20% foetal calf serum. These cell were then plated into a T25 25 flask and placed in a humidified incubator (10% CO₂ in air) at 37°C for 2 days.

After 2 days, the non-attached cells were aspirated. The remaining attached cells were harvested by trypsinisation, and washed two times in DEPC-treated 30 PBS to remove the serum. The pellet was then resuspended into Tri reagent (1 ml) to isolate RNA (Sigma-Aldrich Chemical Co., as described in Sigma Technical Bulletin MB-205). The isolated RNA was quantified by optical density measurements and the absence of contaminating DNA was determined

20

by PCR using β -actin and HPRT primers in separate samples (Wakeman *et al.*, 1998). If free of DNA, the RNA was then used for RT-PCR analysis of Oct4 expression.

5 PCR Amplification of Oct4 from Human EC x Mouse Thymocyte Heterokaryon

In one experiment (2102Ep with thymocytes), a control was prepared, consisting of cells treated as for fusion except that the incubation with PEG 10 was omitted - thus it was anticipated that no 2102Ep x thymocyte heterokaryons would be formed. In another experiment RNA was isolated from thymocytes alone and also from a mouse EC line (PCC4 azal, clone 3), to provide further negative and positive controls for mouse Oct4 expression. cDNA was then produced from the samples using reverse transcriptase (RT) 15 (Wakeman *et al.*, 1998). PCR was then performed using oligonucleotide primers specific for human and mouse *Oct 4*, a marker of pluripotent cells under the standard PCR conditions described in Wakeman *et al.* (1998) with an annealing temperature of 61°C. These products were then subjected to electrophoresis and separated DNA fragments detected by ethidium bromide 20 staining (Figure 7). Molecular size of the amplified fragments was determined by using a 1kb DNA step ladder.

PCR Primers for human and mouse Oct 4

Species	Annealing Temp (°C)	Sequence	Bp	GenBank Accession No. and primer location
<i>Human</i> Forward Reverse	61.4	5'-cgaccatctggcgcttgag-3' 3'-ccccctgtccccattccata-5'	573	X52437 120-139 534-515
<i>Mouse</i> Forward Reverse	60.4	5'-gtccgcggcatacggatgttc-3' 3'-agggggccgcagcttacacat-3'	415	Z11899 361-380 937-918

These primers were designed using the PrimerSelect module of the Lasergene suite of 25 programs (DNAStar Inc., USA). The mouse primers would not be expected to amplify human Oct4.

Enucleation of cells to yield 'cytoplasts' and 'karyoplasts' or 'mini-cells'.

One of the techniques that is employed in our method for producing Re-Programmed Embryonic Stem Cells (RPES cells) is the use of cytochalasin B 5 to generate enucleated EC celis (EC cytoplasts) as the cytoplasm donor, and 'karyoplasts' (also called 'mini-cells') from the differentiated or committed cells as the nucleus donor. Cytochalasin B is well-known to induce cells to extrude their nuclei (Carter, 1967) and has been employed by numerous authors to induce enucleation of a wide range of cells of a variety of species 10 including both mouse and human cells (Poste 1972; Prescott et al 1972; Goldman et al 1973; Wright and Hayflick 1973; Ege and Ringertz 1974a; Wigler and Weinstein 1975). Such enucleation results in a cell lacking a nucleus, but is otherwise intact and viable for a number of days (Goldman et al 1973); these enucleated cells have been called anucleate cells (Poste 1972) or 15 cytoplasts (Veomett et al 1974). The nucleus that is extruded from the cell retains a thin rim of cytoplasm and is surrounded by a plasma membrane; these structures have been called 'karyoplasts' (Veomett et al 1974) or 'mini- cells' (Ege and Ringertz 1975). Enucleation of cells to yield both cytoplasts and karyoplasts may be achieved by well-established techniques in which cells 20 growing attached to a plastic disc are inverted over a solution of cytochalasin B in a centrifuge tube and centrifuged; the cytoplasts remain attached to the plastic disc, while the karyoplasts are pelleted at the bottom of the centrifuge tube (Prescott et al 1972). Alternatively, cells in suspension may be centrifuged through a density gradient, typically composed of Ficoll, 25 containing cytochalasin B (Wigler and Weinstein 1975). In this case, cytoplasts and karyoplasts are formed and may be recovered from different parts of the gradient after centrifugation.

Using the method described by Prescott et al 1972 NTERA-2 clD1 EC cells growing on a plastic disc were inverted over a solution of 7.5 μ g/ml

22

cytochalasin B, in phosphate buffered saline containing 10% fetal bovine serum, in a 50 ml centrifuge tube, and centrifuged for 30 minutes at 12,000 rpm and 37°C in a J2 Bectman Centrifuge using a JA20 rotor. Cytoplasts without nuclei remain attached to the disc. Occasional cells that have escaped 5 enucleation also remain, please see Figure 6. After recovery by incubation overnight in medium without cytochalasin, the discs were fixed in methanol and stained with haematoxylin and cosin (Bar - 50 μ m).

10 Methods for combining (fusing) the cytoplasm of one cell with the nucleus of another.

The methods for creating hybrid cells by fusing two or more cells of different origins together are very well established and widely known. For a review of the commonly used methods based upon Sendai virus induced cell fusion, or 15 cell fusion induced by polyethylene glycol (PEG), see Kennett (1979).

Briefly, mixtures of cells that it is desired to fuse are incubated with a fusogenic agent, such as Sendai virus or PEG, often with centrifugation or agitation to encourage clumping and close apposition of the cell membranes; variables such as time, temperature, cell concentration and fusogenic agent 20 concentration are optimised for each cell combination. An example of cell fusion to produce heterokaryons is presented in Figure 6. Cell fusion was carried out using Polyethylene Glycol 1000, as described by Kennett 1979. Following fusion, the cells were seeded into tissue culture dishes, and incubated in fresh medium overnight. They were then fixed with methanol 25 and stained with haematoxylin and cosin. Several heterokaryons containing 2 and 3 nuclei can be seen in this field. (Bar = 50 μ m).

23

These techniques have also been shown to allow fusion of cytoplasts, prepared by cytochalasin B induced enucleation, with whole cells or karyoplasts, also derived by cytochalasin B induced enucleation (Poste and Reeve 1971; Ege and Ringertz 1975; Ege et al 1973, 1974; Veomett et al 1974; Wright and 5 Hayflick 1975; Shay 1977)).

Another technique that is now well established and widely used for inducing cell fusion, 'electrofusion', involves passing short electric pulses through mixtures of cells (Neil and Zimmermann 1993).

10

Production of RPES cells

The production of RPES cells requires several steps:

1. the selection of appropriate differentiated cells (the Nucleus 15 Donor) and, if necessary, the isolation of their nuclei,
2. the selection of EC cells (the Cytoplasm Donor),
3. the fusion of the differentiated cell nuclei with the EC cells, and
4. the removal of the EC cell nucleus, either before or after fusion.

The production technique may, in some cases, be optimised by pre-treatment 20 of the differentiated cells, or contemporaneous treatment of the differentiated cell/ EC cell fused products, with various agents such as, but not limited to, inhibitors of DNA methylation, to enhance the ability of the differentiated cell nucleus to be re-programmed. After the production of the RPES cells additional methods are required to propagate the cells, to characterise their 25 properties and to induce them to differentiate into required somatic cell types.

Differentiated cells to be used as Nuclear Donors

A large range of somatic cells derived from any tissue or organ of an adult mammal or human, or from embryos or foetuses, or from extra-embryonic tissues such as the trophoblast or yolk sac may be used as a source of nuclei for reprogramming. Particular somatic cell types include but are not limited to thymocytes, peripheral blood lymphocytes, epidermal cells such as from the bucal cavity, cumulus cells, or other stem cells isolated from biopsies of various tissues, such as the bone marrow, the nervous system and the gut. The technique may also be applied to various established cell lines, such as those derived from various tumours including, for example, but not limited to lymphoblastoid cell lines. The selected somatic cells used for the re-programming procedure may be used directly upon isolation or they may be cultured for a short time before further manipulation. In some instances such somatic cells may be combined entirely with EC cells as described below, or nuclei or karyoplasts may first be isolated from them, for example using agents such as cytochalasin B, as discussed above, or by other methods. For example, nuclei may also be isolated using established micromanipulation procedures, or other established cell fractionation procedures.

Parental EC cells to be used as Cytoplasm Donors

20 A large number of EC cell lines have been isolated from human teratocarcinomas, which occur predominantly, but not exclusively, as testicular germ cell tumours. Examples of available human EC cell lines are shown in Table 1. Similarly, EC cell lines derived from teratocarcinomas of the laboratory mouse have also been derived and are readily available.

25 Examples of available mouse EC cell lines are shown in Table 2. To date EC cell lines have not been described from any other species, but if they were derived in the future we would anticipate that they should behave in a manner similar to the existing human and mouse EC cells which resemble one another. Thus, newly isolated EC cells of other species, or from human or mouse

sources, should also be able to re-program differentiated cells to RPES cells as described in its proposal for human and mouse EC cells.

Human EC cells can be readily recognised by a combination of features that include their morphology (see Figure 1), their expression of the cell surface antigens SSEA3 (Shevinsky et al 1982, Andrews et al 1982, 1984b, 1996), SSEA4 (Kannagi et al 1983, Andrews et al 1984b, 1996), and TRA-1-60 and TRA-1-81 (Andrews et al 1984a, 1984b, 1996), and typically by low expression or absence of SSEA1 (Andrews et al 1980, 1982, 1984b, 1996) (see Figure 2). By contrast, mouse EC cells typically express SSEA1 (Solter and Knowles, 1978) but not SSEA3 (Shevinsky et al 1982), SSEA4 (Kannagi et al 1983) or TRA-1-60 or TRA-1-81 (Andrews et al 1984a). Human EC cells like mouse EC cells express high levels of alkaline phosphatase (ALP) (Bernstine et al 1973, Benham et al 1981); in the case of human EC cells most ALP activity is due to expression of the liver/bone/kidney isoform which can be detected as a cell surface antigen by monoclonal antibodies TRA-2-59 and TRA-2-54 (Andrews et al 1984c). In common with ES cells and primordial germ cells, EC cells also typically express the transcription factor Oct3/4 (Rosfjord and Rizzino 1994; Brehm et al 1998).

20 Fusion of parental differentiated cells and parental EC cells to yield RPES cells

Several methods may be used to combine the cytoplasm of an EC cell and the nucleus of a differentiated cell to yield an RPES containing the nuclear genome of the differentiated cell but not the EC cell.

25 A. Cells may be fused by use of chemical agents such as polyethylene glycol (PEG) or viruses such as Sendai virus, or by passing an electric current through a mixture of cells. As discussed above, these methods

26

are well known and may be readily applied. These methods may be used to fuse:

1. a differentiated cell with an EC cell, or
2. a karyoplast from a differentiated cell with an EC cell, or
- 5 3. a differentiated cell with one or more cytoplasts isolated from EC cells, or
4. a karyoplast from a differentiated cell with one or more cytoplasts isolated from EC cells.

10 In cases (1) and (2), the result will initially be a heterokaryon containing two nuclei, one from each parental cell. If this heterokaryon were allowed to divide the result would be a hybrid cell containing a single nucleus with a complete or partial genome from each parental cell. However, in our method of producing RPES cells, the EC nucleus is removed prior to cell division of the hybrid cell, so that the derivative 15 dividing cell population retains only the genome of the parental differentiated cell.

20 In cases (3) and (4) the EC nucleus is removed from the EC cell before fusion, for example by enucleation with cytochalasin B as discussed above, so that the resulting product contains only the differentiated cell nucleus and cytoplasm from the EC cell parent. In any of these cases, the resulting RPES cells that continue to proliferate retain only the nuclear genome of the differentiated parental cell, which is now reprogrammed to express a new pattern of gene activity.

25 In cases (1) and (2) the EC cell nucleus is removed from the heterokaryon in one of several ways that include, but are not limited to, partial enucleation using drugs such as cytochalasin B, applied in the same manner as described above for enucleating EC cells and

generating cytoplasts for fusion. In the present case in which enucleation is carried out after fusion, some heterokaryons lose both nuclei, in which case they do not proliferate, some heterokaryons lose the differentiated cell nucleus, in which case they retain the parental EC nucleus and continue proliferating, some heterokaryons lose the EC cell nucleus, in which case they continue proliferating as RPES cells, and some heterokaryons retain both nuclei and eventually continue proliferating as hybrid cells. Several methods are used to select the RPES cells and to eliminate any of the cells retaining an EC cell genome or to eliminate any cells retaining a somatic nucleus that has failed to undergo re-programming. In one method, the proliferating cells are cloned by established techniques (e.g. by picking single cells with a micropipette - see Andrews et al 1982, 1984b), and individual clones are screened using genetic markers for those that retain an EC genome. The latter cells are discarded, whereas those that retain only a differentiated cell genome but not an EC cell derived genome, and express an RPES phenotype, are retained. Standard DNA genotyping techniques using well established DNA fingerprinting technology (Jeffreys et al 1985, 1988; Yan et al 1996) may be used to identify whether the nuclear genome of any proliferating cells is derived from either the EC cell or differentiated cell parent, or both.

In another method, before use as a fusion partner, the EC cell parent is genetically marked by insertion of a gene that will allow selection against any cell carrying that gene; for example, the EC cell can be stably transfected with a vector encoding the Herpes Simplex Virus-1 Tk gene (HSV1-Tk), such that any cells carrying that gene can be killed by culture in the presence of a number of drugs including acyclovir (9-[(2-hydroxyethoxy)methyl]guanine) or FIAU (1-(2-deoxy-2-fluoro- β -D-arabinofuranosyl)-5-iodouracil) (Borrelli et al 1988; Hasty et al

1991), or gancyclovir (Rubinstein et al 1993; McCarrick and Andrews 1992). In this method, following partial enucleation, the remaining heterokaryons are cultured in medium containing this drug, and only those that have lost the EC cell nucleus survive. Other selectable 5 genetic systems can also be similarly used. Persisting parental differentiated cells that have not been reprogrammed are removed by cloning the surviving cells, or by selecting RPES cells by virtue of their expression of specific surface antigen markers that include, but are not limited to, SSEA3, SSEA4, TRA-1-60 or TRA-1-81, as discussed 10 above as characteristic markers of EC cells. These same markers have been shown to be expressed by ES cells derived directly from human embryos (Thomson et al 1998; Shambrott et al 1998). For the latter approach, fluorescence activated cell sorting (FACS), a widely used 15 method for separating subsets of cells can be used (e.g. Andrews et al 1982, 1987; Ackerman et al 1994; Williams et al 1988).

In another method, the EC cell parent is incubated prior to fusion, with 20 a drug that irreversibly inactivates its nucleus and prevents its replication, for example, topoisomerase inhibitors such as etoposide (Downes et al 1991; Fulka and Moor 1993). The resulting heterokaryon naturally eliminates this treated nucleus prior to cell division, so that the resulting dividing cell population only contains the genome derived from the parental differentiated cell. This approach 25 may also be combined with the preceding 'partial' enucleation of heterokaryons' approach to ensure complete loss of the EC genome.

In another method, after cell fusion to produce a heterokaryon, the EC cell nucleus is removed by micro-manipulation.

B. Rather than chemical, viral or electrically induced fusion, the nucleus of the differentiated cell is combined with an EC cell parent by micro-manipulation. In this method, the nucleus of the differentiated

cell is withdrawn using a micropipette inserted through the cell membrane. It is then injected either into an inoculated EC cell, or into an intact EC. In the later case the EC cell nucleus is then removed by a similar technique, or by one of the techniques described above, before 5 nuclear fusion and cell division occurs.

Growth and selection of RPES cells

Following fusion to combine a differentiated cell and an EC cells, with prior or subsequent removal of the EC cell nucleus, it is necessary to provide 10 appropriate conditions for the re-programming of the differentiated cell nucleus and for the subsequent proliferation of the resulting RPES cells.

Several methods are used to enhance the efficiency of reprogramming:

1. prior to fusion the differentiated cell and EC cell are synchronised with respect to position in the cell cycle, by use of reversible 15 inhibitors that arrest the cell cycle at specific stages (e.g. nocodazole), or by the use of conditions such as low serum to arrest cells in G1, or by selection of cells at specific stages of the cell cycle by using vital DNA stains and flow microfluorimetry (Fluorescence Activated Cell Sorting) (Ashihara and Baserga 1979; Andrews et al 1987; Crissman 1995; Stein et al 1995).
2. the differentiated cell or the immediate fusion product is cultured in the presence of drugs that inhibit methylation or promote demethylation (e.g. 5-azacytidine) (e.g. Taylor and Jones 1979; Jones 1985; Keshet et al 1986), or alter the structure of chromatin, 25 for example butyrate, spermine, trichostatin A or trapoxin which inhibit deacetylation and promote acetylation of histones, which plays a role in X chromosome inactivation, gene imprinting and

30

regulation of gene expression (Caldarera et al 1975; McKnight et al 1980; Stein et al 1997; Hu et al 1998; Wolffe and Pruss 1996;).

5 3. the period of time between production of heterokaryons and the removal of the EC cell nucleus is made as long as possible without permitting nuclear fusion. This period can be elongated by culturing the heterokaryons under conditions that reversibly inhibit progress through the cell cycle (e.g. thymidine block - Stein et al 1995), or by altering growth conditions, such as serum starvation or lowered temperature, that retard cell division but permit reprogramming to proceed.

10 4. any, or all combinations of these methods.

In all these experiments the cells are cultured in standard cell culture media that include but are not restricted to Dulbecco's modified Eagle's Medium (DME, high glucose formulation) or Ham's F12, supplemented in some cases with foetal bovine serum or with other additives (e.g. see Andrews et al 1980, 1982, 1984, 1994). Subsequent to fusion and re-programming, the growth of the resulting cells may be optimised culture on feeder layers of cells that include, but are not restricted to, irradiated or mitomycin C treated STO cells, or embryonic fibroblasts of various species, including humans (see Robertson 1987a; Thomson et al 1998). The cells may be cultured in the presence of various growth factors or other tissue culture additives, that include but are not restricted to LIF, FGF, SCF.

Differentiation of the RPES cells

25 In the best cases, the RPES cells acquire pluripotent properties that closely resemble those of embryonic stem cells, so that the RPES cells are able to differentiate and to initiate differentiation pathways that result in the formation of any cell type that may be found in the adult, embryo or in extra-embryonic

tissues, given appropriate conditions. The maintenance of an EC cell state can be monitored by assay of various markers that include the cell surface antigens SSEA3, SSEA4, TRA-1-60, TRA-1-81, by their expression of alkaline phosphatase and by expression of Oct3/4, as discussed above. The RPES cells 5 typically retain their stem cell phenotype when cultured on appropriate feeder cells. However, they can initiate differentiation under a variety of circumstances.

Thus removal from feeder cells, or culture in suspension, followed by 10 replating in the absence of feeder cells in appropriate tissue culture flasks results in differentiation of stem cells into a variety of cell types that include neurons, muscle of various sorts and haematopoietic cells (see descriptions in Robertson 1987a). Differentiation of pluripotent stem cells (e.g. see Figures 3 and 4) may also be initiated by altered conditions affecting cell density and 15 aggregation (e.g. seeding at low cell densities or trypsinisation) or exposure to various agents that include but are not restricted to retinoic acid, and other retinoids, hexamethylene bisacetamide, and the bone morphogenetic proteins (see Robertson 1987a; Andrews 1984; Andrews et al 1982, 1990, 1994, 1996; Thomson et al 1998). The type of cells that arise depend upon the nature of 20 the inducing agent, and the culture conditions including the presence or absence of specific growth factors or other molecules.

Discussion

25 Although pluripotent stem cell lines have been derived from early embryos (Robertson, 1987b; Thomson et al 1995, 1998), primordial germ cells (Matsui et al 1992; Shambrott et al 1998) and from germ cell tumours (reviewed, Andrews, 1998) of various species, including the laboratory mouse, rhesus monkeys and humans, and nuclei from differentiated somatic adult cells have

been re-programmed to yield embryonic stem cells by transplantation to enucleated oocytes (Campbell *et al* 1996; Wakayama *et al* 1998), there are no reports that pluripotent stem cells, resembling embryonic stem cells with the capacity to differentiate into a variety of functional somatic cell types, can be 5 produced by the re-programming of differentiated or committed embryonic or adult somatic cells, or extra-embryonic cells, without the use of oocytes.

We now describe methods by which embryonal carcinoma (EC) cells, derived 10 from teratocarcinomas, can be used to re-program various somatic, differentiated cells, or other embryonic or extra-embryonic cell types, to a state from which they can then be induced to differentiate into one or more functional differentiated cell types that are distinct from the parental cells. In 15 the best cases, but not necessarily in all cases, the re-programmed cells produced by this technique, called 'Re-programmed Embryonic Stem Cells' (RPES cells), resemble embryonic stem cells derived directly from early embryos, and can be induced to differentiate into a broad range of functional, differentiated cell types that include, but are not limited to, neurons, muscle (including skeletal and cardiac muscle) and haematopoietic cells. These RPES cells are diploid with a normal karyotype, and isogenic with the differentiated 20 parental cells from which they are derived. They may be used to generate differentiated cells for transplantation and use in cell and tissue replacement therapies.

In some cases, only partial reprogramming occurs with, for example, the activation of several genes that are not active in the parental differentiated 25 nuclear donor cell. Such cells are also of use in a variety of these same circumstances.

An example of such a gene is Oct4. Oct4 has previously been reported to be characteristically expressed by undifferentiated EC and ES cells (Brehm *et al.*,

33

1998). Therefore, to test the ability of human EC cell cytoplasm to reprogram somatic cells, isolated mouse thymocytes were fused with human EC cells, (2102Ep, clone 4D3 (Andrews *et al.*, 1982) or TERA1 (Fogh and Trempe, 1975; Andrews *et al.*, 1980)), to produce heterokaryons which were tested 5 after 2 days for activation of Oct4 expression from the thymocyte genome. Evidence for such activation would indicate, not only that human EC cells are capable of re-programming a somatic cell nucleus to an ES/EC cell like state, but also that the regulatory factors involved are capable of working between different mammalian species. Thus if human EC cells can reprogram a mouse 10 somatic cell, we would anticipate not only that they would be able to reprogram a human somatic cell, but also that mouse EC cells would be able to reprogram human somatic cells as well. Similarly, given the resemblance of EC and ES cells, it would be expected that ES cells could reprogram somatic cells in the same way as EC cells.

15

In Experiment 1, as anticipated, an amplified band (573 bp), corresponding to human Oct4 expression was detected similarly in RNA preparations from the 2102Ep x thymocyte fusion in the presence of PEG, and in the mock fusion in the absence of PEG, consistent with its expression by 2102Ep human EC cells. 20 However, a band corresponding to mouse Oct4 (415 bp) was only detected in the RNA preparation from the 2102Ep x thymocyte fusion in the presence of PEG, when heterokaryons were expected to be present. The corresponding absence of mouse Oct4 from the mock fusion indicates both the absence of Oct4 expression from mouse thymocytes in this experiment, and the 25 requirement for formation of heterokaryons for its activation from the thymocyte genome by the 2102Ep cytoplasm. No products were seen in the 'water' control, indicating absence of contamination.

30 In a second experiment, in which 2102Ep and TERA1 human EC cells were fused with mouse thymocytes in the presence of PEG, mouse Oct4 was only

detected in the 2102Ep fusion, again confirming the ability of 2102Ep cells to reprogram mouse thymocytes with activation of Oct4 expression, but suggesting in this experiment that TERA1 cytoplasm did not achieve reprogramming. In both cases, human Oct4 was detected as expected, 5 consistent with its expression by 2102Ep or TERA1 human EC cells.

In further controls, no mouse Oct4 expression was detected in RNA prepared from isolated mouse thymocytes not used for fusion. However, a similar sized PCR band to that detected in the 2102Ep x thymocyte fusion samples, corresponding to mouse Oct4, was detected in mouse PCC4 EC cells as 10 expected.

In our method, RPES cells are created by combining the nucleus from a differentiated or committed cell (the Nuclear donor), whether from adults or from embryos, with the cytoplasm from an EC cell (the Cytoplasm donor), 15 from which the nucleus is removed. Several methods can be used to combine the nucleus from the differentiated cell and the cytoplasm from the EC cell; in some methods the EC cell nucleus is removed prior to combination of the cytoplasm with the donated nucleus, and in other methods the EC cell nucleus is removed after combination. If EC cells and differentiated cells from the 20 same species are used, then the resulting RPES cells retain cytoplasmic genetic determinants (e.g. the mitochondrial genome) and a nuclear genome from the same species. By contrast, embryonic stem-like cells produced by transplantation of somatic cells into enucleated oocytes of other species will continue to harbour mitochondria of that other species. Especially for the 25 production of human RPES cells and their differentiated derivatives for transplantation into a human host, the maintenance of a human nuclear and human cytoplasmic genome could be a distinct advantage. Further, RPES cells that are isogenic with the anticipated human host can be produced by this technique without resort to any embryo, so avoiding practical difficulties that 30 may be associated, for example, with immune rejection upon transplantation to

the human host, and also obviating ethical difficulties inherent in the use of human embryos.

The method that we describe incorporates the techniques for maintaining and propagating the RPES cells produced, and the techniques for inducing them to 5 differentiate into a range of differentiated, functional cell types.

References

10 Andrews P.W. and Goodfellow P.N. (1980) Antigen expression by somatic cell hybrids of a murine embryonal carcinoma cell with thymocytes and L cells. *S0000omat. Cell Genet.* 6: 271-284.

15 Andrews P.W., Bronson D.L., Benham F., Strickland S. and Knowles B.B. (1980) A comparative study of eight cell lines derived from human testicular teratocarcinoma. *Int. J. Cancer* 26: 269-280.

20 Andrews P.W., Goodfellow P.N., Shevinsky L., Bronson D. L. and Knowles B.B. (1982) Cell surface antigens of a clonal human embryonal carcinoma cell line: Morphological and antigenic differentiation in culture. *Int. J. Cancer* 29: 523-531.

25 Andrews P.W. (1982) Human embryonal carcinoma cells in culture do not synthesize fibronectin until they differentiate. *Int. J. Cancer* 30: 567-571.

Andrews P.W., Damjanov I., Simon D., Banting G., Carlin C., Dracopoli N.C. and Fogh J. (1984b) Pluripotent embryonal carcinoma clones derived from the human teratocarcinoma cell line Tera-2: Differentiation *in vivo* and *in vitro*. *Lab. Invest.* 50: 147-162.

30 Andrews P.W., Meyer L.J., Bednarz K.L. and Harris H. (1984c) Two monoclonal antibodies recognizing determinants on human embryonal

carcinoma cells react specifically with the liver isozyme of human alkaline phosphatase. *Hybridoma* 3: 33-39.

Andrews P.W. (1984) Retinoic acid induces neuronal differentiation of a cloned human embryonal carcinoma cell line in vitro. *Dev. Biol.* 103: 285-293.

Andrews P.W., Banting G.S., Damjanov I., Arnaud D. and Avner P. (1984a) Three monoclonal antibodies defining distinct differentiation antigens associated with different high molecular weight polypeptides on the surface of human embryonal carcinoma cells. *Hybridoma* 3: 347-361.

10 Andrews P.W., Gönczöl E., Plotkin S.A., Dignazio M. and Oosterhuis J.W. (1986) Differentiation of TERA-2 human embryonal carcinoma cells into neurons and HCMV permissive cells: Induction by agents other than retinoic acid. *Differentiation* 31: 119-126.

15 Andrews, P.W., Oosterhuis J.W. and Damjanov I. (1987) Cell lines from human germ cell lines. In: *Teratocarcinomas and embryonic stem cells: A practical approach* (E.J. Robertson, ed.). IRL Press, Oxford, pp 207-248.

20 Andrews P.A., Nudelman E., Hakomori S. -i. and Fenderson B.A. (1990). Different patterns of glycolipid antigens are expressed following differentiation of TERA-2 human embryonal carcinoma cells induced by retinoic acid, hexamethylene bisacetamide (HMBA) or bromodeoxyuridine (BUdR). *Differentiation* 43, 131-138.

Andrews P.W., Damjanov I., Berends J., Kumpf S., Zappavigna V. Mavilio F. and Sampath K. (1994). Inhibition of proliferation and induction of differentiation of pluripotent human embryonal carcinoma cells by osteogenic protein-1 (or bone morphogenetic protein-7). *Laboratory Investigation*, 71, 243-251.

25 Ashihara, J., and Basergu, R. (1979). Cell Synchronisation Methods. *Enzymology*, 58, 248-262.

Bernstine, E.F., Hooper, M.L., Grandchamp, S., Ephrussi, B. (1973). Alkaline phosphatase activity in mouse teratoma. *Proc Natl Acad Sci USA* 70, 3899-3902.

Brehm, A., Ovitt, C.E., Schöler, H.R. (1978). Oct 4: more than just a
5 POUerful marker of the mammalian germline? *Acta Path. Mirobiol, Immunol, Scand.* 106, 114-126.

Bronson, D.L., Andrews, P.W., Solter, D., Cervenka, J., Lange, P.H., and Fraley, E.E. (1980). A cell line derived from a metastasis of a human testicular germ-cell tumour. *Cancer Res.* 40, 2500-2506.

Bronson, D.L., Ritzi, D.M., Fraley, E.E., and Dalton, A.J. (1978). Morphologic evidence for retrovirus production by epithelial cells derived from a human testicular tumour metastasis. *J.Natl. Cancer Inst.* 60, 1305-1308.

15 Bronson, D.L., Andrews, P.W., Vessella, R.L., and Fraley, E.E. (1983). In vitro differentiation of human embryonal carcinoma cells. In "Teratocarcinoma Stem Cells" (L.M. Silver, G.R. Martin and S. Strickland, eds.), *Cold Spring Harbor Conferences on Cell Proliferation*, Vol. 10, pp. 597-605. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York.

Bronson, D.L., Saxinger, W.C., Ritzi, D.M., Fraley, E.E. (1984). Production of virions with retrovirus morphology by human embryonal carcinoma cells in vitro. *J. Gen. Virol* 65, 1043-1051.

25 Campbell, K.H.S., McWhir, J., Ritchje, W.A., Wilmut, I. (1996). Sheep cloned by nuclear transfer from a cultured cell line. *Nature*, 380, 64-66.

Carter, S.B. (1967). Effects of cytochalasins on mammalian cells. *Nature*, 213, 261-264.

Downes, C.S., Mullinger, A.M., Johnson, R.T. (1991). Inhibitions of DNA topoisomerase II prevent sistem chromatid separation in mammalian cells but do not prevent exit from mitosis. *Proc. Nat. Acad. Sci (USA)*. 88, 8895-8899.

5

Ege, T., and Ringertz, N.R., (1974). Preparation of microcells by enucleation of micronucleate cells. *Exp. Cells Res.* 87, 378-382.

10 Ege, T., Krondahl, U., Ringertz, N.R. (1974). Introduction of nuclei and micronuclei into cells and enucleated cytoplasms by Sendai virus induced fusion. *Ex.p Cell Res.* 88, 428-432.

15 Ege, T., and Ringertz, N.R. (1975). Viability of cells reconstituted by virus-induced fusion of mini cells with enucleate cells. *Expl. Cell Res.*, 94, 469-473.

Ege, T., Zeuthen, J., Ringertz, N.R. (1973). Cell fusion with enucleated cytoplasms. *Nobel*, 23, 189-194.

20 Engström, W., Tally, M., Granerus, M., Hedley, E.P., and Schofield, P. (1991). Growth factors and the control of human teratoma cell proliferation. *Recent Res. Cancer Res.* 123, 145-153.

25 Fenderson, B.A., Andrews, P.W., Nudelman, E., Clausen, H., and Hakomori, S.-I., (1987). Glycolipid core structure switching from clobo- to lacto- and ganglio-series during retinoic acid-induced differentiation of TERA-2 derived human embryonal carcinoma cells. *Dev. Biol.* 122, 21-34

Fogh, J., and Trempe, G. (1975). New human tumor cell lines. In "Human Tumor Cells in vitro" (J.Fogh, ed.), pp. 115-159. Plenum Press, New York.

30

Fulka, J., Moor, R.M. (1993). Non invasive chemical enucleation of mouse oocytes. *Mol. Reprod. Devel.*, 34, 427-430.

5 Goldman, R.D., Pillack, R., Hopkins, N.H. (1973). Preservation of normal behaviour by enucleated cells in culture. *Proc. Nat. Acad. Sci. (USA)*, 70, 750-754.

10 Gönczol, E., Andrews, P.W., and Plotkin, S.A. (1984). Cytomegalovirus replicates in differentiated but not undifferentiated human embryonal carcinoma cells. *Science* 224, 159-161.

Gurdon J. *The control of gene expression in animal development* (Oxford University Press 1974).

15 Hogan, B., Fellous, M., Avner, P., and Jacob, F. (1977). Isolation of a human teratoma cell line which expresses F9 antigen. *Nature (London)* 270, 515-518.

20 Jakob, H., Boon, T., Galliard, J., Nicolas, J.F., Jacob, F. (1973). Térotocarcinome de la souris: isolement, culture et propriétés de cellules à potentialités multiples. *Ann Microbiol (Paris)* 124b 269-282.

Jeffreys, A.J., Wilson, V., Thein, S.L. (1985). Individual specific fingerprints of human DNA. *Nature*, 316, 76-79

25 Jeffreys, A.J., Wilson, V., Newmann, R., Keyte, J. (1988). Amplification of human minisatellites by the polymerase chain reaction - towards DNA fingerprinting of single cells.

30 Kennett, R.H. (1979). Cell Fusion in: *Cell Culture, Methods in Enzymology*. (eds. Jakoby, W.B., and Pastan, I.H.) Academic Press San Diego, 58, 345-359.

Matsui, Y., Szebo, K., and Hogan, B.L.M. (1992). Derivation of pluripotential embryonic stem cells for murine primordial germ cells in culture. *Cell*, 70, 841-847.

5

Mavilio F., Simeone A., Boncinelli E. and Andrews P.W. (1988) Activation of four homeobox gene clusters in human embryonal carcinoma cells induced to differentiate by retinoic acid. *Differentiation*, 37,73-79.

10 Miller., W.H., and Dmitrovsky, E. (1991). Growth factors in human germ cell cancer. *Recent Res.Cancer Res.* 123, 183-189.

Neil, G.A., and Simmermann, U. (1993). Electrofusion, Methods in Enzymology, 220, 174-196.

15 Poste, G., and Reeve, P. (1971). Formation of hybrid cells and heterokaryous by fusion of enucleated and nucleated cells. *Nature New Biology*, 229, 122-125.

20 Poste, G. (1972). Enucleation of mammalian cells by cytochalasin B. *Exp. Cell Res.* 73, 273-286.

Prescott, D.M., Myerson, D., Wallace, J. (1972). Enucleation of mammalian cells with cytochalasin B. *exp. Cell Res.* 71, 480-485.

25 Robertson, E.J. (Editor) (1987). *Teratocarcinomas and embryonic stem cells: A practical approach*. IRL Press, Oxford.

Robertson, E.J., (1987b). Embryo-derived stem cell lines in (Robertson, E.J., Editor) *Teratocarcinomas and embryonic stem cells: A practical approach*. IRL Press, Oxford, pp 71-112.

5 Rosfjord, E., and Rizzino A. (1994). The octaner motif present in the Rex-1 promoter binds Oct-1 and Oct-3 expressed by EC cells and ES cells. *Biochem. Biophys. Res. Counc.*, 203, 1795-1803.

10 Shambrott, M.J., Axelman, J., Wang, S., Bugg, E.M., Littlefield, J.W., Donovan, P.J., Blumenthal, P.D., Huggins, G.R., Gearhart, J.D. (1998). Derivation of pluripotent stem cells from cultured human primordial germ cells. *Proc. Nat. Acad. Sci. USA*, 95, 13726-13731.

15 Shay, J.W. (1997). Selection of recontributed cells from karyoplasts fused to chloramphenical resistant cytoplasts. *Proc. Nat. Acad. Sci. USA*. 74, 2461-2464.

20 Shevinsky, L.H., Knowles, B.B., Damjanov, I., Solter, D. (1982). Monclonal antibody to murine embryo defines a stage specific embryonic antigen expression on mouse embryos and human teratocarcinoma cells. *Cell* 30, 697-705.

25 Simeone, A., Acampora, D., Arcioni, L., Andrews, P.W., Böncinelli, E., and Mavilio, F. (1990). Sequential activation of human HOX2 homeobox genes by retinoic acid in human embryonal carcinoma cells. *Nature (London)* 346, 763-766.

30 Solter, D., and Knowles, B.B. (1978). Monoclonal antibody defining a stage specific mouse embryonic antigen (SSEA-1). *Proc Natl Acad Sci (USA)*. 75, 5565-5569.

Teshima, S., Shimosato, Y., Hirohashi, S., Tome, Y., Hayashi, I., Kanazawa, H., and Kakizoe, T., (1988). Four new human germ cell tumor cell lines. *Lab. Invest.* 59, 328-336.

5

Thomson, J.A., Itskovitz-Eldor, J., Shapira, S.S., Waknitz, K.A., Swiergiel, J.J., Marshall, V.S., Jones, J.M. (1998). Embryonic stem cell lines derived from human blastocysts. *Science*, 282, 1145-1147.

10 Thompson, S., Stern, P.L., Webb, M., Walsh, F.S., Engström, W., Evans, E.P., Shi, W.K., Hopkins, B., and Graham, F.F. (1984). Cloned human teratoma cells differentiate into neuron-like cells and other cell types in retinoic acid. *J. Cell Sci.* 72, 37-64.

15 Veomett, G., Prescott, D.M., Shay, J., Porter, K.R. (1974). Reconstruction of mammalian cells from nuclear and cytoplasmic components separated by treatment with cytocholasin B. *Proc Nat Acad Sci*, 71, 1999-2002.

20 Vogelzang, N., Andrews, P.W., and Bronson, D. (1983). An extragonadal human embryonal carcinoma cell line 1618K. *Proc. Am. Assoc. Cancer Res.* 24, 3.

25 Von Keitz, A.T., Riedmiller, H., Neuman, K., Grutschart, W., Fonatsch, C. (1994). Establishment and characterisation of a seminoma cell line (S2). *Invest. Urol (Berlin)*, 5, 28-31.

Wakayama, T., Perry, A.C.F., Succotti, M., Johnson, K.R., Yanagimachi, R. (1998). Full term development of mice from enucleated oocytes injected with cumulus cell nuclei. *Nature*, 394, 369-374.

Wakeman, J.A., Heath, P.R., Pearson, R.C.A., Andrews, P.W. (1997) MAL mRNA is induced during the differentiation of human embryonal carcinoma cells into neurons, and is also localised within specific regions of the human brain. *Differentiation* 62:97-105.

5 Wakeman, J.A., Walsh, J., Andrews, P.W., (1998). Human Wnt-13 is developmentally regulated during the differentiation of NTERA-2 pluripotent human embryonal carcinoma cells. *Oncogene* 17:179-186

10 Wang, N., Trend, B., Bronson, D.L., and Fraley, E.E. (1980). Nonrandom abnormalities in chromosome 1 in human testicular cancers. *Cancer Res.* 40, 796-802.

15 Wang, N., Perkins, K.L., Bronson, D.L., and Fraley, E.E. (1981). Cytogenetic evidence for pre-meiotic transformation of human testicular cancers. *Cancer Res.* 41, 2135-2140.

20 Wenk, J., Andrews, P.W., Casper, J., Hata, J-I., Pera, M.F., von Keitz, A., Damjanov, I., Fenderson, B.A. 1994. Glycolipids of germ cell tumours: extended globo-series glycolipids are a hallmark of human embryonal carcinoma cells. *Int. J. Cancer.*, 58, 108-115.

Wigler, M.H., and Weinstein, I.B. (1975). A preparative method for obtaining enucleated mammalian cells. *Biochem Biophys Res Comm.* 63, 669-674.

25 Williams B.P., Daniels G.L., Pym B., Sheer D., Povey S., Okubo Y., Andrews P.W. and Goodfellow P.N. (1988) Biochemical and genetic analysis of the OKa blood group antigen. *Immunogenetics*, 27, 322-329.

44

Wright, W.E., Hayflick, L. (1973). Enucleation of cultured human cells. Proc. Soc. Exp. Biol. and Med. 144, 587-592.

Wright, W.E., and Hayflick L. (1975). Use of biochemical lesions for 5 selection of human cells with hybrid cytoplasms. Proc. Nat. Acad. Sci (USA). 72, 1812-1816.

Yan, R., Ottenbreit, K., Hukku, B., Mally, M., Chou, S.R., Kaplan, J. (1996). DNA fingerprinting of human cell lines using PCR amplification of fragment 10 length polymorphisms. In Vitro Cellular and Developmental Biology, 32, 656-662.

Table 1
Human EC Cell Cytoplasmic Donors (this list is illustrative but not comprehensive)

Cell Line	Phenotype in Culture ^{1,2}	Surface Antigen Expression ^{1,3}						Tumour Origin			Xenograft	References
		SSEA-3	SSEA-4	TRA-1-60	TRA-1-81	TRA-2-40	Original Site	Biopsy Site	Mitology			
1218B	EC	+	+	+	+	TRA-2-54 [L-ALP] ⁺	Testis	Primary	EC,S	EC	Andrews <i>et al</i> 1980 Wang <i>et al</i> 1980, 1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994	Andrews <i>et al</i> 1980 Wang <i>et al</i> 1980, 1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
833KE	EC	+	+	+	+		Testis	Primary	EC,T,C,S	EC	Bronson <i>et al</i> 1978, 1980, 1984	Bronson <i>et al</i> 1978, 1980, 1984
TERA1	EC	+	+	+	+		Testis	Lung metastasis	EC,T		Wang <i>et al</i> 1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994	Wang <i>et al</i> 1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
SuS1	EC	+	+	+	+		Testis		EC,T		Hogen <i>et al</i> 1977 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994	Hogen <i>et al</i> 1977 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
NCCIT	EC	+	+	+	+		Testis		EC,T		Teshima <i>et al</i> 1988 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994	Teshima <i>et al</i> 1988 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
1618K	EC	+	+	+	+		Retinoblastoma	Primary	EC,T,Y	EC,T,C,Y		
1777N Rpmel	EC	+	+	+	+		Retinoblastoma		BL		Vogelzang <i>et al</i> 1983 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994	Vogelzang <i>et al</i> 1983 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
82	7 not EC	-	-	-	-		Testis	Retropertitoneal Lymph node	EC		Bronson <i>et al</i> 1983, 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994	Bronson <i>et al</i> 1983, 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
							Testis	Primary	SCID Germinal centers	7EC	Von Keltz 1994 Wenk <i>et al</i> 1994 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994	Von Keltz 1994 Wenk <i>et al</i> 1994 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994

Table 1
Human EC Cell Cytoplasmic Donors (this list is illustrative but not comprehensive)

Cell Line	Phenotype In Culture ^{1,2}	Surface Antigen Expression ^{1,3}					Tumour Origin			Xenograft	References
		SSEA3	SSEA4	TRA1-60	TRA1-81	TRA2-49 (L-ALP)*	Original Site	Biopsy Site	Histology		
TERA2	EC	+	+	+	+	+	Testis	Lung Metastasis	EC,T	Pugh & Trempe 1975 Wang <i>et al</i> 1981 Andrews <i>et al</i> 1984 Gündüz <i>et al</i> 1984	
Including sublines such as NTERA-2 and clones										Andrews <i>et al</i> 1984	
										Thompson <i>et al</i> 1984 Pendergrass <i>et al</i> 1987 Simeone <i>et al</i> 1990 Engstrom <i>et al</i> 1991 Miller & Dmitrovsky 1991 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994	
Z102Ep	EC	+	+	+	+	+	Testis	Primary	EC,T,Y	EC	Andrews <i>et al</i> 1980, 1982 Wang <i>et al</i> 1980,1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
Z102R Rpm1	EC	+	+	+	+	+	Testis	Retropertitoneal lymphnodes	EC,T	Wang <i>et al</i> 1980,1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994	
1156QF	EC	+	+	+	+	+	Testis	Primary	EC,C,S	Andrews <i>et al</i> 1980 Wang <i>et al</i> 1980,1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994	

Table 1
Human EC Cell Cytoplasmic Donors (this list is illustrative but not comprehensive)

Notes

- 1 The phenotype of cells in culture is based upon observations of morphology, growth patterns and antigen expression (see, Andrews *et al* 1996).
- 2 Culture conditions: Human EC cells can generally be maintained in Dulbecco's Modified Eagles medium (DMEM), high glucose formulation, supplemented with glutamine and 10% foetal bovine serum, but other media have also been used (e.g. RPMI for NCCIT) (see Andrews and Danjanov 1994). High cell densities (75×10^6 per 75 cm² tissue culture flask) are optimal for maintaining an EC phenotype (see Andrews *et al* 1982, 1984b, Andrews and Danjanov 1994). All the cell lines may be harvested for passage using 0.25% trypsin and 2mM EDTA, in Ca²⁺/Mg²⁺ - free Dulbecco's phosphate buffered saline, but in some cases (TERA-2 and derivatives, and SuSa) clumping of cells is preferable for best maintenance of an EC phenotype. In the latter cases, cells are harvested for passage by scraping, for example with glass beads, rather than by use of trypsin (see Andrews *et al* 1984b, Andrews and Danjanov 1994).
- 3 Surface antigen expression: SSEA-3, SSEA-4, TRA-1-60 and TRA-1-81 expression is characteristic of human EC cell lines but the level of expression of these antigens is variable and appears to reflect their state of differentiation (e.g. see Andrews *et al* 1996, Andrews *et al* 1982, Kannagi *et al* 1983, Andrews *et al* 1984a, Shevinsky *et al* 1982).
- 4 High levels of the liver/bone/kidney isoforms of alkaline phosphatase (L-ALP) detected as a cell surface antigen by monoclonal antibodies TRA-2-49 and TRA-2-54 are also characteristic of human EC cells (Benham *et al* 1981, Andrews *et al* 1984c, Andrews *et al* 1996).
- 5 Differentiation and loss of EC phenotype: Many human EC cells undergo morphological changes and change in surface antigen expression, notably the induction of SSEA-1 and down regulation of SSEA-3, when cultured at low cell densities and well dispersed (Andrews *et al* 1982, 1984b). These changes appear to represent a limited capacity for differentiation. Other lines differentiate extensively if exposed to agents such as retinoic acid (e.g. NTERA-2, Andrews 1984; NCCIT, Teshima *et al* 1988), hexamethylene bisacetamide (e.g. NTERA-2, Andrews *et al* 1986, 1990), and the bone morphogenetic proteins (e.g. NTERA-2, Andrews *et al* 1994). Differentiation in the latter cases is marked by loss of the characteristic EC marker antigens, appearance of new antigens (e.g. see, Fenderson *et al* 1987), activation of new genes (e.g. Hox genes, Mavilo *et al* 1988; Mai, Wakeman *et al* 1997; Whi-13, Wakeman *et al* 1998).

Table 2
Mouse EC Cell Cytoplasmic Donors (this list is illustrative but not comprehensive)

Cell Line	Surface Antigen Expression SSEA-1 ¹	Reference
F9	+	Bernstine <i>et al</i> 1993, Solter and Knowles 1978
PCC4	+	Jakob <i>et al</i> 1973, Solter and Knowles 1978
PCC3 (ND1)	+	Jakob <i>et al</i> 1973, Solter and Knowles 1978
MH-15	+	Solter and Knowles 1978
FA-25	+	Solter and Knowles 1978

¹ Solter and Knowles 1978

CLAIMS

1. A cell comprising at least part of the cytoplasm derived from an embryonal teratocarcinoma cell combined with a nucleus of a somatic cell.

5

2. A cell according to Claim 1 wherein said cell is a cybrid characterised by the possession of at least one pluripotential characteristic.

3. A cell according to Claim 2 characterised in that said pluripotential 10 characteristic is the ability to differentiate into at least one selected tissue type.

4. A cell according to Claim 2 characterised in that said pluripotential characteristic includes the ability of said cell to proliferate in culture in an undifferentiated state.

15

5. A cell according to Claim 4 characterised in that said cell has the capacity to proliferate in continuous culture in an undifferentiated state for at least 6 months and ideally 12 months.

20 6. A cell according to any of Claims 2-5 characterised in that said pluripotential characteristic includes the expression of at least one selected marker.

7. A cell according to Claim 6 characterised in that said pluripotential characteristic is expression of Oct4.

25

8. A cell according to Claim 6 characterised in that said selected marker is a cell surface marker.

30 9. A cell according to Claim 8 characterised in that said cell surface marker is selected from the group including SSEA-1 (-); and/or SSEA-3 (+); and/or SSEA-4 (+); and/or TRA-1-60 (+); and/or TRA-1-81 (+); and/or alkaline phosphatase (+).

10 10. A cell according to any of Claims 2-9 characterised in that said pluripotential characteristic includes the presence of telomerase activity.

5 11. A cell according to any of Claims 2-10 characterised in that said pluripotential characteristic includes the presence of a chromosomal methylation pattern characteristic of pluripotential cells.

10 12. A cell according to any of Claims 2-11 characterised in that said pluripotential characteristic includes the ability to induce tumours when introduced into an animal.

13. A cell-line comprising cells according to any of Claims 1-12.

15 14. A cell-line according to Claim 12 characterised in that said cell-line is of human origin.

15 15. A method for the preparation of a cytoplasmic part for use in the production of a cell according to any of Claims 1-12 or a cell-line according to Claims 13 or 14 comprising;

25 (i) providing at least one embryonal teratocarcinoma cell;
(ii) separating at least part of the cytoplasm from the nucleus of said cell;
(iii) isolating said cytoplasmic part; and, optionally
(iv) storing said isolated cytoplasmic part prior to use.

16. A method according to Claim 15 characterised in that said cytoplasmic part is a cytoplasm.

30 17. A method for preparing a cell according to any of Claims 1-12 or a cell-line according to Claims 13 or 14 comprising;

- (i) combining at least one embryonal teratocarcinoma cell with at least one somatic cell;
- (ii) removing the embryonal teratocarcinoma nucleus from said combined cell;
- (iii) culturing said cell under conditions conducive to proliferation and expansion of said cell; and, optionally
- (iv) storing said cell culture under suitable conditions.

10 18. A method of combining at least part of the cytoplasm of an embryonal teratocarcinoma cell with a somatic cell comprising;

- (i) providing at least part of the cytoplasm of an embryonal teratocarcinoma cell;
- (ii) combining said cytoplasmic part with at least one somatic cell;
- (iii) growing said combined cell in culture; and, optionally
- (iv) storing said combined cell under suitable storage conditions.

15 19. A method according to Claim 18 characterised in that said cytoplasmic part is provided as a cytoplasm.

20 20. A method according to Claims 18 or 19 characterised in that said cytoplasm is combined with said somatic cell via cytoplasm/somatic cell fusion.

25 21. A method according to any of Claims 18-20 characterised in that said embryonal carcinoma cell and said somatic cell are of human origin.

22. A cell culture comprising at least one cell according to the invention.

30 23. A method for inducing differentiation of at least one cell according to Claims 1-12 comprising:

- (i) providing a cell according to any of Claims 1-12;
- (ii) culturing said cell under conditions conducive to the differentiation of said cell into at least one tissue; and, optionally
- 5 (iii) storage of said differentiated tissue prior to use under suitable storage conditions.

24. A method according to Claim 23 characterised in that said culture conditions are selected so as to provide a tissue type selected from at least one of: neural, 10 smooth muscle, striated muscle, cardiac muscle, bone, cartilage, liver, kidney, respiratory epithelium, haematopoietic cells, spleen, skin, stomach, intestine.

25. At least one tissue type or organ comprising at least one cell according to any of Claims 1-12.

15

26. A therapeutic composition comprising at least one cell according to any of Claims 1-12 and a suitable excipient, diluant or carrier.

27. A therapeutic composition according to Claim 26 for use in tissue 20 transplantation.

28. A method to treat conditions or diseases requiring transplantation of tissue comprising:

- 25 (i) providing at least one tissue type or organ according to the invention;
- (ii) surgically introducing said tissue type or organ to a patient to be treated; and
- (iii) treating said patient under conditions which are conducive to the acceptance of said transplanted tissue by said patient.

30

29. A kit comprising at least one cell according to the invention; instructions with respect to the maintenance of said cell in culture; and, optionally, factors required to induce differentiation of said cell to at least one desired tissue type or organ.

5

10

15

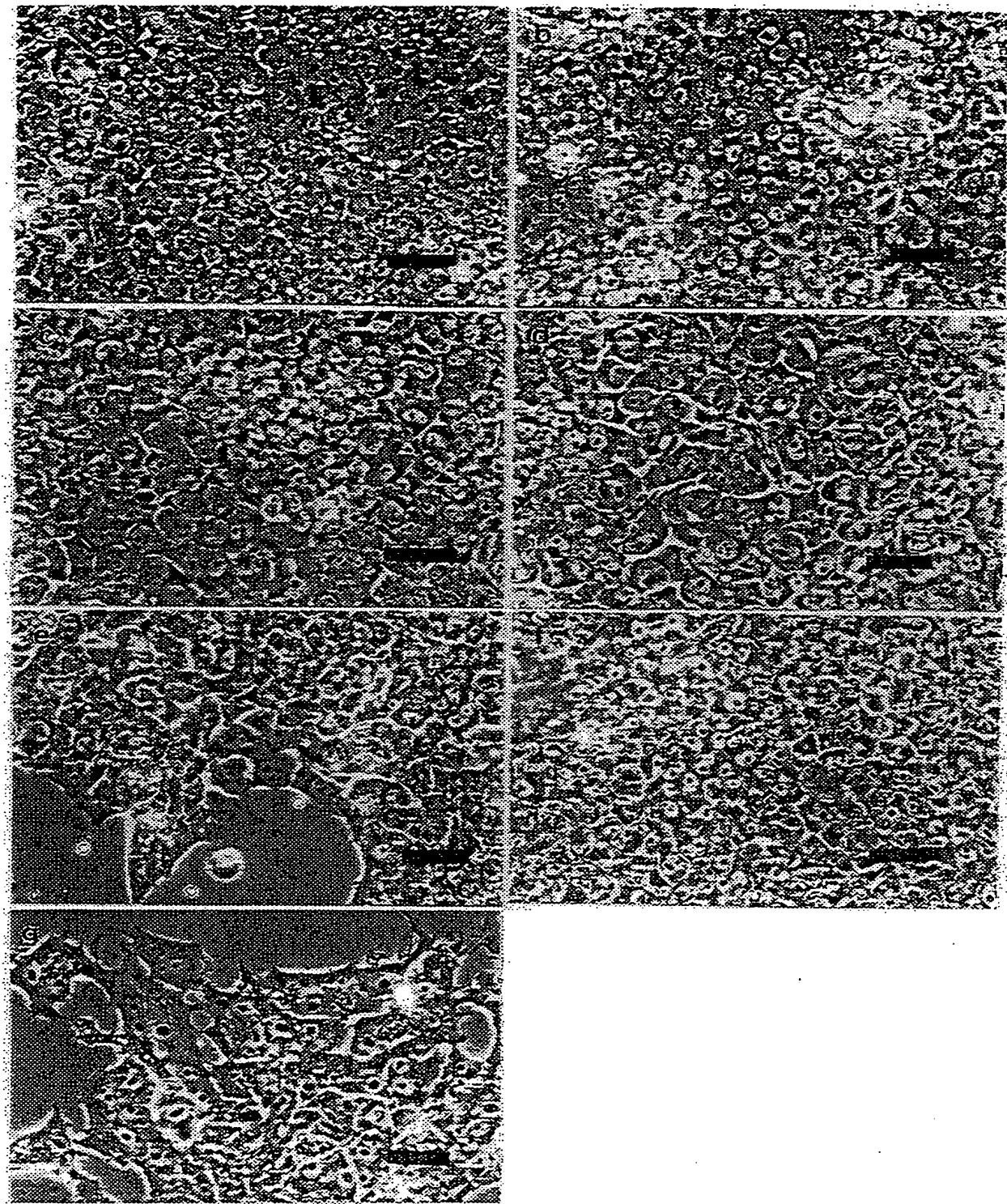
20

25

30

THIS PAGE BLANK (USPTO)

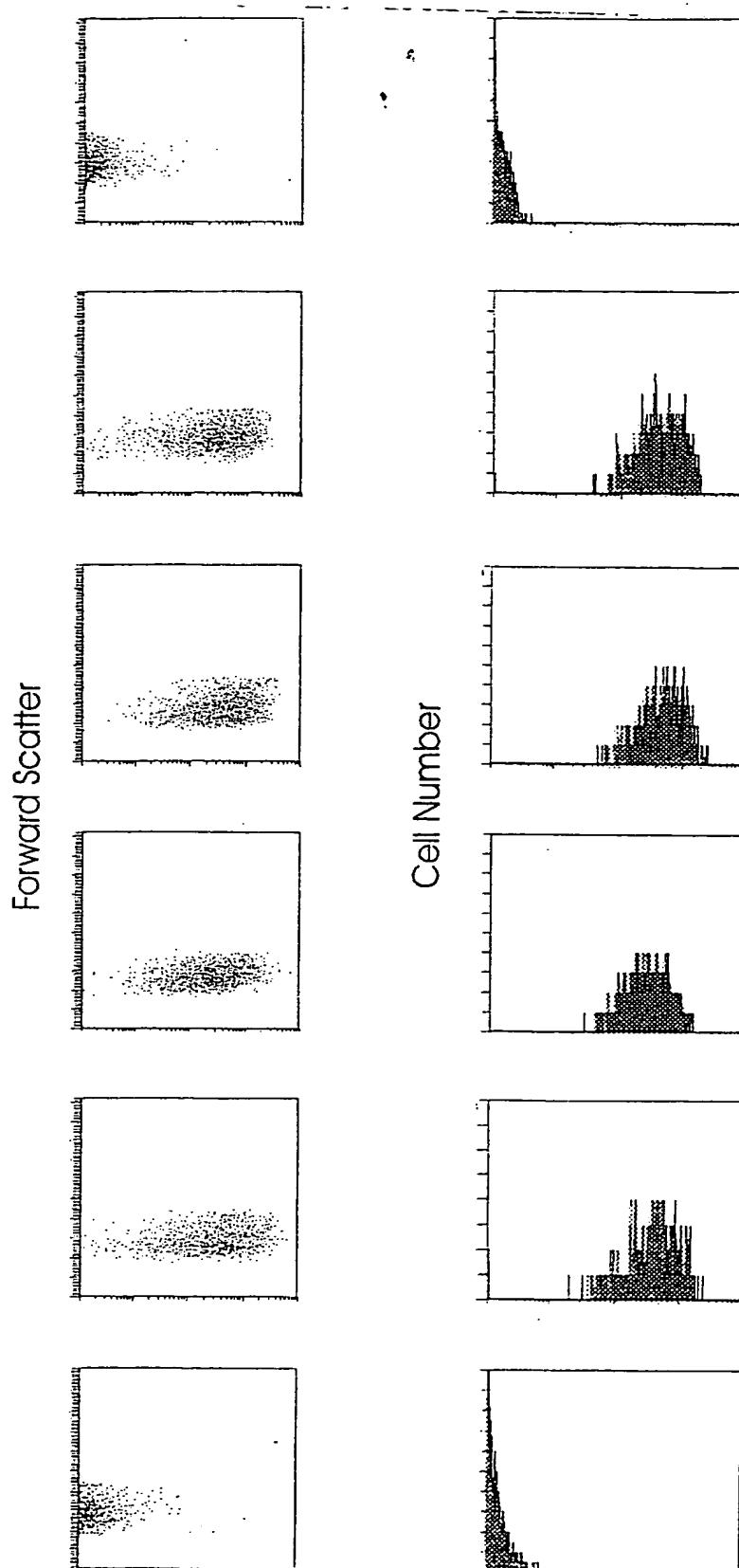
FIGURE 1



513 Rec'd PCT/PTO 20 AUG 2001

THIS PAGE BLANK (USPTO)

NO PAGE BLANK (USPTO)



513 Rec'd PCT/PTO 20 AUG 2001

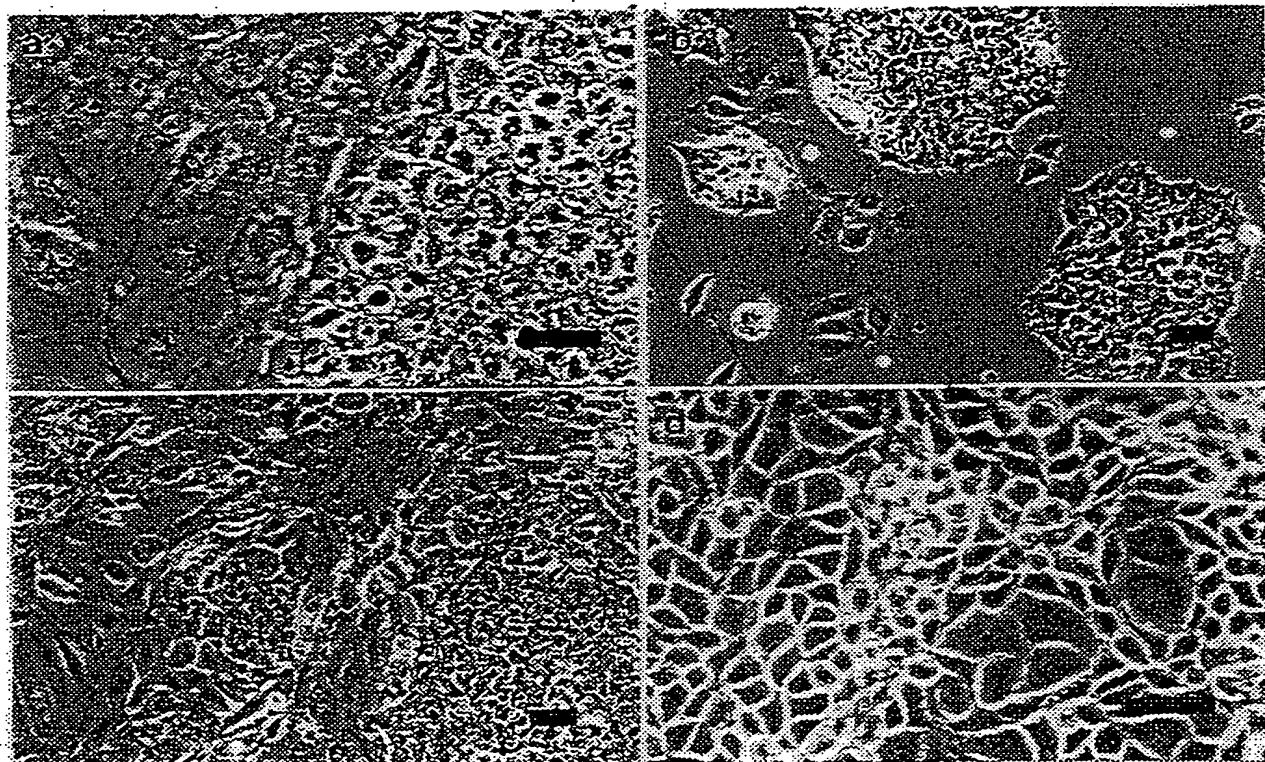
THIS PAGE BLANK (USPTO)

WO 00/49138

Title: PLURIPOTENTIAL
CELLS-1
Inventor(s): Peter Andrews
DOCKET NO.: 033236-0116

09/913853
PCT/GB00/00582

FIGURE 3



SUBSTITUTE SHEET (RULE 26)

513 Rec'd PCT/PTO 20 AUG 2001

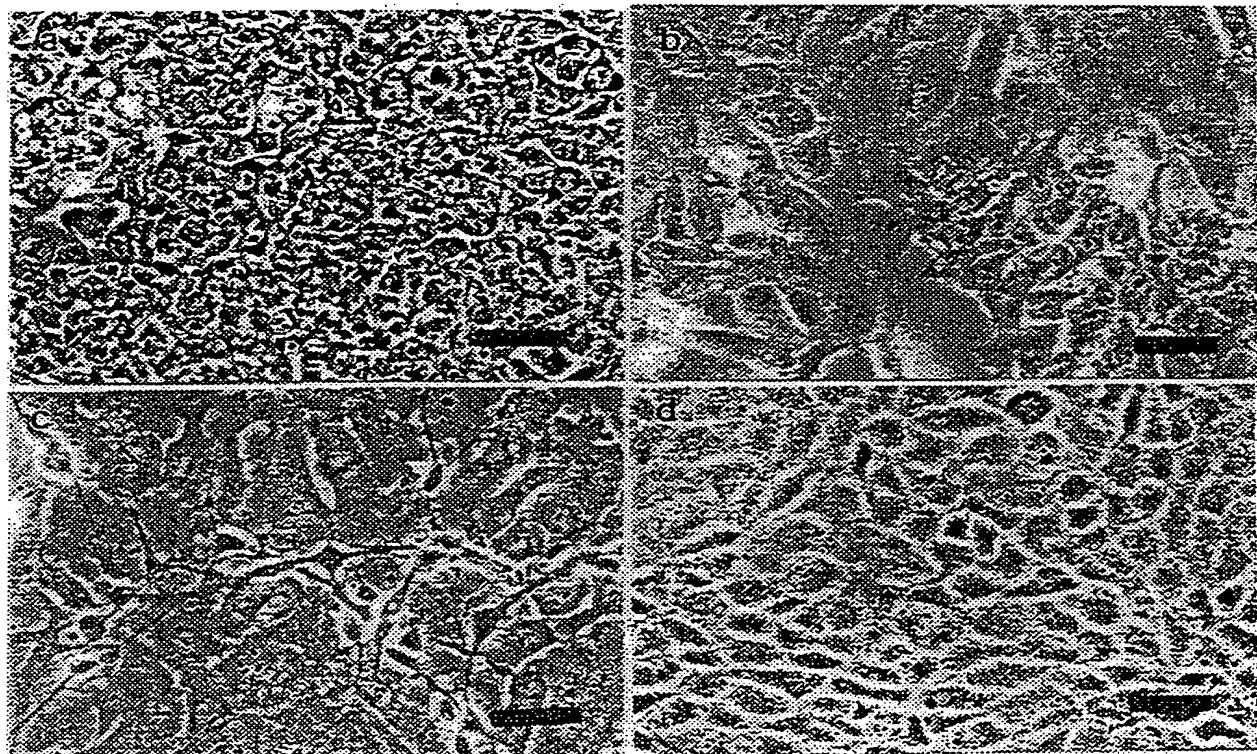
THIS PAGE BLANK (USPTO)

WO 00/49138

Title: PLURIPOTENTIAL
CELLS-1
Inventor(s): Peter Andrews
DOCKET NO.: 033236-0116

09/913853
PCT/GB00/00582

FIGURE 4



518 Rec'd PCT/PTO 20 AUG 2001

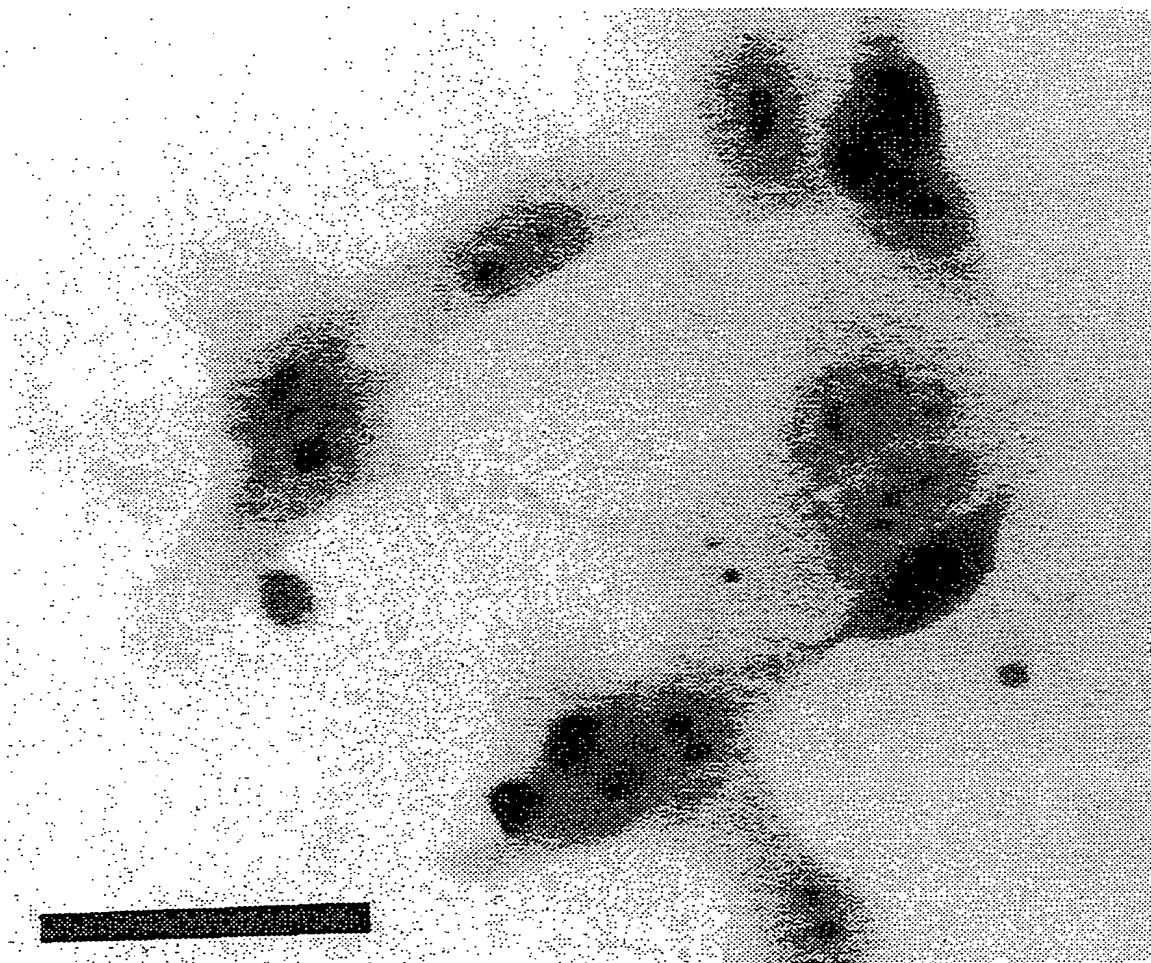
THIS PAGE BLANK (USPTO)

WO 00/49138

Title: PLURIPOTENTIAL
CELLS-1
Inventor(s): Peter Andrews
DOCKET NO.: 033236-0116

09/913853
PCT/GB00/00582

FIGURE 5



SUBSTITUTE SHEET (RULE 26)

518 Rec'd PCT/PTO 20 AUG 2001

THIS PAGE BLANK (USPTO)

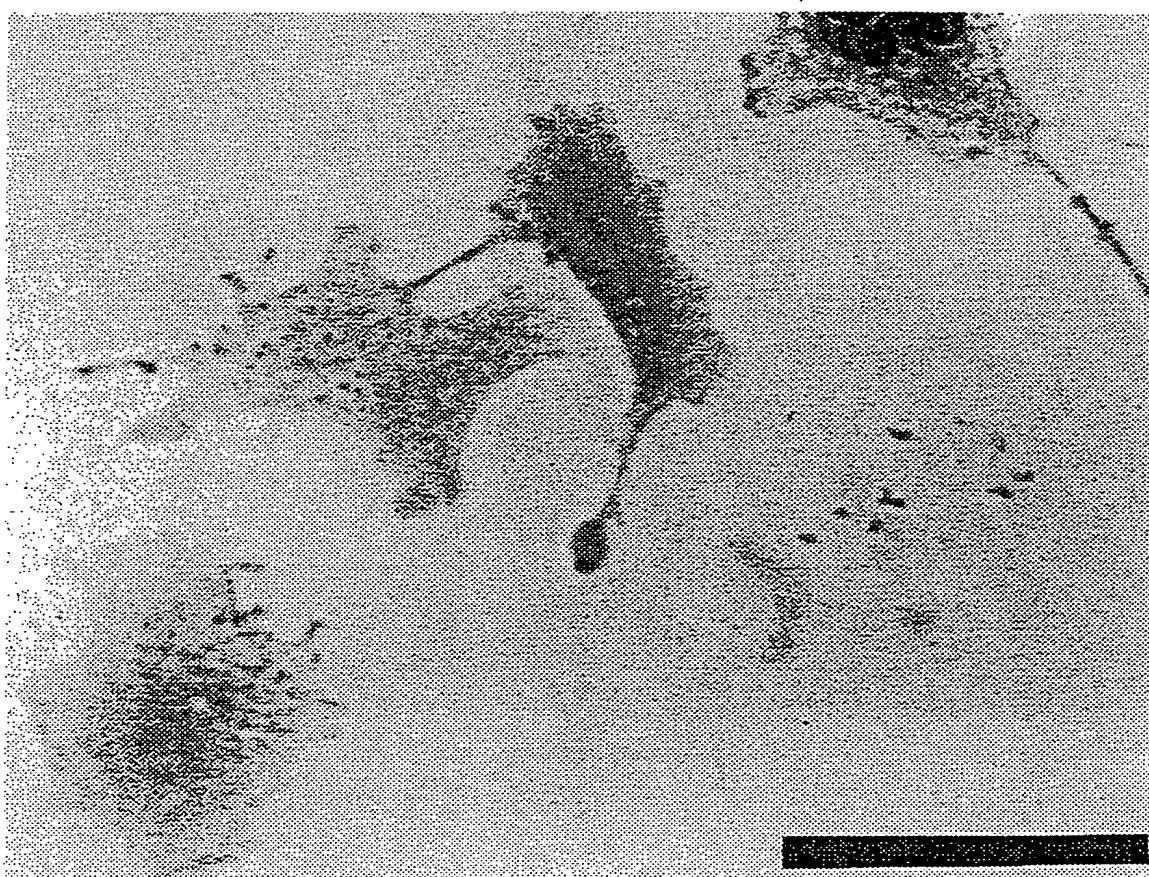
WO 00/49138

Title: PLURIPOTENTIAL
CELLS-1
Inventor(s): Peter Andrews
DOCKET NO.: 033236-0116

09/913853

PCT/GB00/00582

FIGURE 6

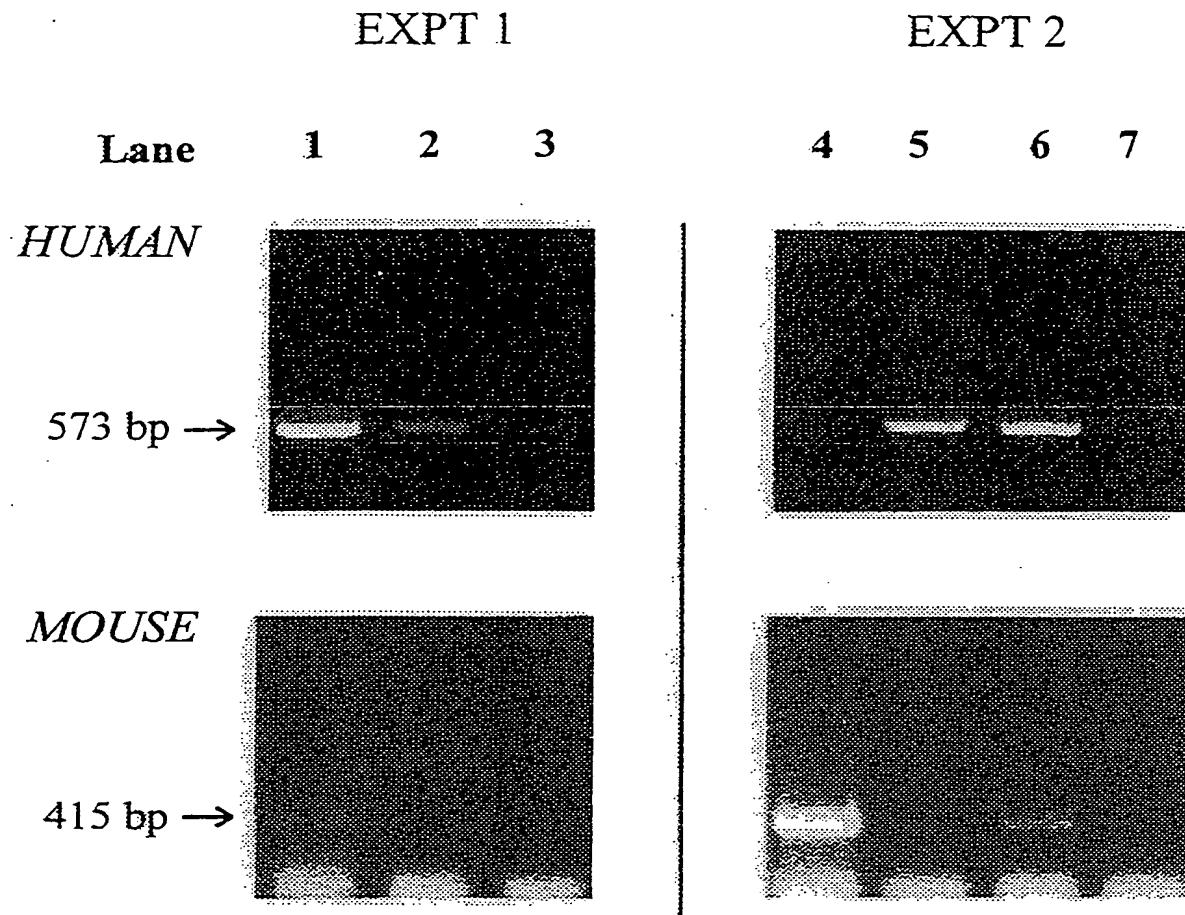


SUBSTITUTE SHEET (RULE 26)

518 Rec'd PCT/PTO 20 AUG 2001

THIS PAGE BLANK (USPTO)

FIGURE 7



Lane

- 1 2102Ep (2×10^6) x Thymocytes (2×10^7) with PEG FUSION
- 2 2102Ep (2×10^6) x Thymocytes (2×10^7) with NO FUSION
- 3 Water
- 4 PCC4 cells (3×10^6)
- 5 TERA1 (3×10^6) x Thymocytes (3×10^7) with PEG FUSION
- 6 2102Ep (3×10^6) x Thymocytes (3×10^7) with PEG FUSION
- 7 Thymocytes (3×10^6)

518 Rec'd PCT/US 20 AUG 2001

THIS PAGE BLANK (USPTO)

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
24 August 2000 (24.08.2000)

PCT

(10) International Publication Number
WO 00/49138 A3

(51) International Patent Classification⁷: C12N 5/22, A61K 35/12, 35/54, A61P 43/00 // C12N 5/28

(21) International Application Number: PCT/GB00/00582

(22) International Filing Date: 18 February 2000 (18.02.2000)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
9903805.1 20 February 1999 (20.02.1999) GB

(71) Applicant (for all designated States except US): UNIVERSITY OF SHEFFIELD [GB/GB]; Western Bank, Sheffield S10 2TN (GB).

(72) Inventors; and

(75) Inventors/Applicants (for US only): ANDREWS, Peter [GB/GB]; University of Sheffield, Dept. of Biomedical Sciences, Wester Bank, Sheffield S10 2TN (GB). KEMP, Paul [GB/GB]; 16 Chadkirk Road, Romiley SK6 3JY (GB).

(74) Agent: HARRISON GODDARD FOOTE; Tower House, Merrion Way, Leeds LS2 8PA (GB).

(81) Designated States (national): AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

— With international search report.

(88) Date of publication of the international search report:
22 February 2001

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

WO 00/49138 A3

(54) Title: PLURIPOTENTIAL CELLS DERIVED FROM EMBRYONAL TERATOCARCINOMA CELLS AND THE NUCLEUS OF STOMATIC CELLS

(57) Abstract: The invention relates to isolated pluripotential cells comprising at least part of the cytoplasm from a teratocarcinoma cell and a nucleus of a somatic cell. The invention also relates to methods to prepare such cells.

THIS PAGE BLANK (USPTO)

PENT COOPERATION TREATY

PCT

INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference RCD/P38040WO	FOR FURTHER ACTION see Notification of Transmittal of International Search Report (Form PCT/ISA/220) as well as, where applicable, item 5 below.	
International application No. PCT/GB 00/ 00582	International filing date (day/month/year) 18/02/2000	(Earliest) Priority Date (day/month/year) 20/02/1999
Applicant UNIVERSITY OF SHEFFIELD et al.		

This International Search Report has been prepared by this International Searching Authority and is transmitted to the applicant according to Article 18. A copy is being transmitted to the International Bureau.

This International Search Report consists of a total of 4 sheets.

It is also accompanied by a copy of each prior art document cited in this report.

1. Basis of the report

a. With regard to the language, the international search was carried out on the basis of the international application in the language in which it was filed, unless otherwise indicated under this item.

the international search was carried out on the basis of a translation of the international application furnished to this Authority (Rule 23.1(b)).

b. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of the sequence listing :

contained in the international application in written form.

filed together with the international application in computer readable form.

furnished subsequently to this Authority in written form.

furnished subsequently to this Authority in computer readable form.

the statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.

the statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished

2. Certain claims were found unsearchable (See Box I).3. Unity of Invention is lacking (see Box II).

4. With regard to the title,

the text is approved as submitted by the applicant.

the text has been established by this Authority to read as follows:

PLURIPOENTIAL CELLS DERIVED FROM EMBRYONAL TERATOCARCINOMA CELLS AND THE NUCLEUS OF STOMATIC CELLS

5. With regard to the abstract,

the text is approved as submitted by the applicant.

the text has been established, according to Rule 38.2(b), by this Authority as it appears in Box III. The applicant may, within one month from the date of mailing of this international search report, submit comments to this Authority.

6. The figure of the drawings to be published with the abstract is Figure No.

as suggested by the applicant.

because the applicant failed to suggest a figure.

because this figure better characterizes the invention.

None of the figures.

THIS PAGE BLANK (USPTO)

INTERNATIONAL SEARCH REPORT

International Application No

PCT/00/00582

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N5/22 A61K35/12 A61K35/54 A61P43/00 //C12N5/28

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ, BIOSIS, MEDLINE, EMBASE, CHEM ABS Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>WANGH LAWRENCE J ET AL: "Efficient reactivation of Xenopus erythrocyte nuclei in Xenopus egg extracts." JOURNAL OF CELL SCIENCE, vol. 108, no. 6, 1995, pages 2187-2196, XP000939319 ISSN: 0021-9533 the whole document</p> <p>---</p> <p>-/-</p>	1-3

 Further documents are listed in the continuation of box C. Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

Date of mailing of the international search report

12 September 2000

25/09/2000

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl.
Fax: (+31-70) 340-3016

Authorized officer

Le Flao, K

THIS PAGE BLANK (USPTO)

INTERNATIONAL SEARCH REPORT

International Application No

PCT/GB 00/00582

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	TOSU M ET AL: "CLONAL ISOLATION AND CHARACTERIZATION OF MYOBLAST-LIKE RECONSTITUTED CELLS FORMED BY FUSION OF KARYOPLASTS FROM MOUSE TERATOCARCINOMA CELLS WITH RAT MYOBLAST CYTOPLASTS" CELL STRUCTURE AND FUNCTION, vol. 13, no. 3, 1988, pages 249-266, XP000939352 ISSN: 0386-7196 abstract	1-3
A	J A THOMSON ET AL: "EMBRYONIC STEM CELL LINES DERIVED FROM HUMAN BLASTOCYSTS" SCIENCE, AMERICAN ASSOCIATION FOR THE ADVANCEMENT OF SCIENCE, US, vol. 282, 6 November 1998 (1998-11-06), pages 1145-1147, XP002121340 ISSN: 0036-8075 the whole document	1-29
A	YEOM ET AL: "Germline regulatory element of Oct-4 specific for the totipotent cycle of embryonal cells" DEVELOPMENT, GB, COLCHESTER, ESSEX, vol. 122, 1996, pages 881-894, XP002094592 ISSN: 0950-1991 abstract	7
A	WO 97 07669 A (ROSLIN INSTITUTE) 6 March 1997 (1997-03-06) claims 1-19	1-29
A	FULKA J ET AL: "CLONING BY SOMATIC CELL NUCLEAR TRANSFER" MICROBIOLOGICAL REVIEWS, US, AMERICAN SOCIETY FOR MICROBIOLOGY, WASHINGTON, DC, vol. 20, 1998, pages 847-851, XP002900701 ISSN: 0146-0749 the whole document	1-29

THIS PAGE BLANK (USPTO)

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/GB 00/00582

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9707669	A 06-03-1997	AU 716956 B	09-03-2000
		AU 6831096 A	19-03-1997
		BR 9610034 A	21-12-1999
		CA 2229568 A	06-03-1997
		CN 1202084 A	16-12-1998
		CZ 9800608 A	15-07-1998
		EP 0849990 A	01-07-1998
		EP 0930009 A	21-07-1999
		EP 1005789 A	07-06-2000
		GB 2318578 A,B	29-04-1998
		GB 2331751 A,B	02-06-1999
		HU 9900234 A	28-05-1999
		JP 2000506722 T	06-06-2000
		NO 980845 A	29-04-1998
		NZ 316149 A	28-10-1999
		PL 325331 A	20-07-1998

THIS PAGE BLANK (USPTO)